

3. Stem Cells

Potential and Niches

- Additional (optional) textbooks:
 Michal K Stachowiak & Emmanuel S Tzanakakis, “ *Stem Cells: From Mechanisms to Technologies*” - World Scientific, 2011. <http://www.worldscientific.com/>

STEM CELLS

From Mechanisms to Technologies

Advances in stem cell biology and biotechnology have sparked hopes that therapies will soon be available for maladies which were considered incurable before. However, realization of the clinical potential of stem cells will require better understanding of stem cell physiology and the development of advanced technologies for their efficient differentiation in medically relevant quantities. Resolving these issues necessitates synergistic approaches from multiple fields. Systems biology can be employed to dissect the mechanisms regulating the genome and proteome of stem cells during self-renewal and commitment. Microfluidic platforms can be used to recreate aspects of the stem cell niche and obtain a better understanding of the interactions among stem cells and with their environment. The milieu of stem cells and their progeny can be shaped with appropriately designed biomaterials for the engineering of tissues to replace, reconstitute or regenerate damaged organs. To that end, enabling bioreactor technologies will be necessary for the generation of large quantities of stem cells and their derivatives in a robust and cost-efficient manner. This book invites world-renowned experts in the above fields to discuss the latest advances in their respective areas and to provide insights on the future challenges and achievements in the area of stem cells.

STEM CELLS

From Mechanisms to Technologies

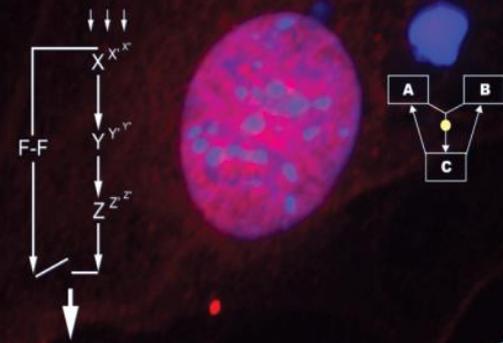
Stachowiak
Tzanakakis

STEM CELLS

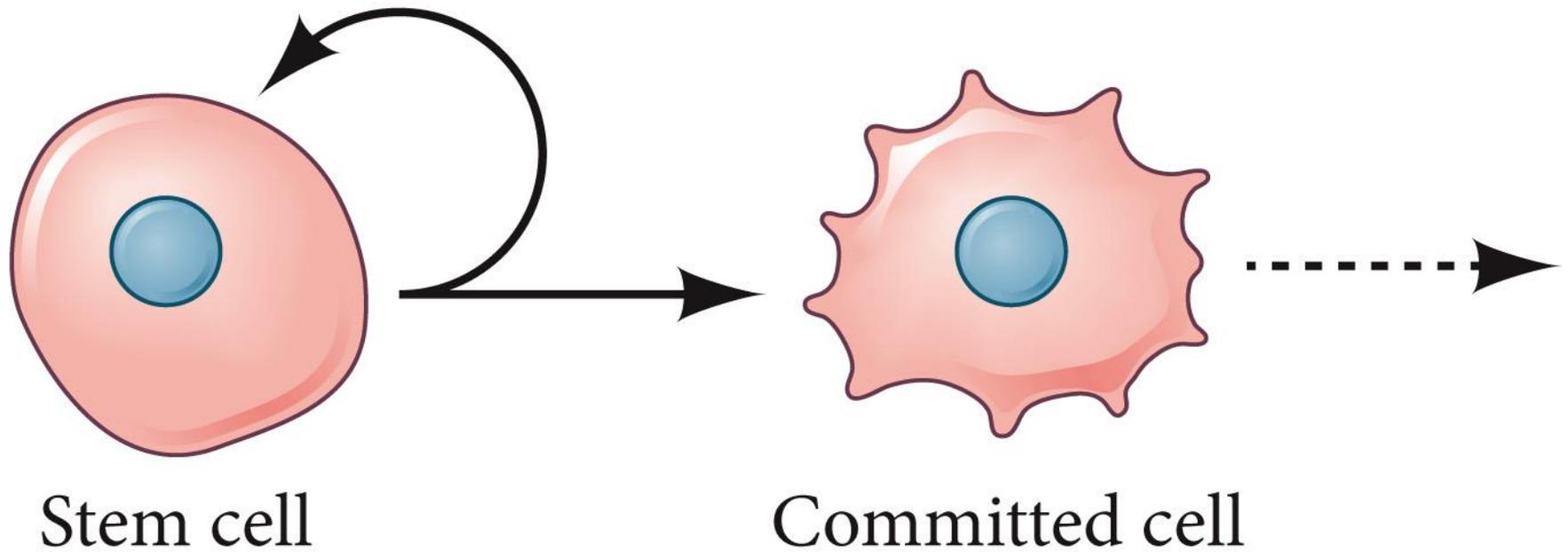
From Mechanisms to Technologies

Michal K Stachowiak
Emmanuel S Tzanakakis

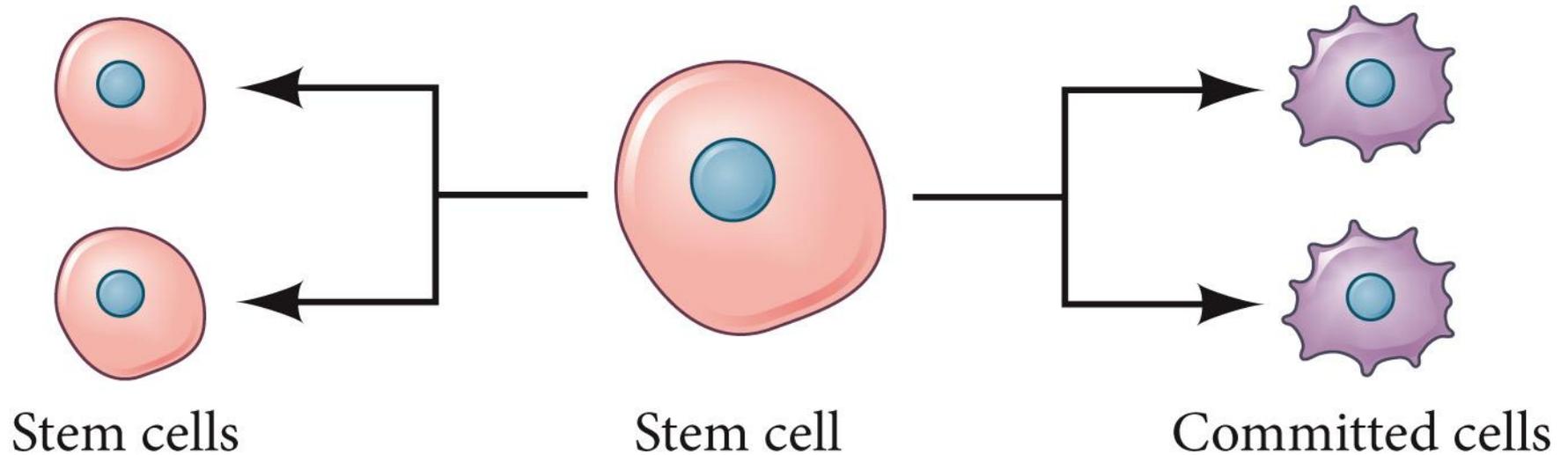
editors



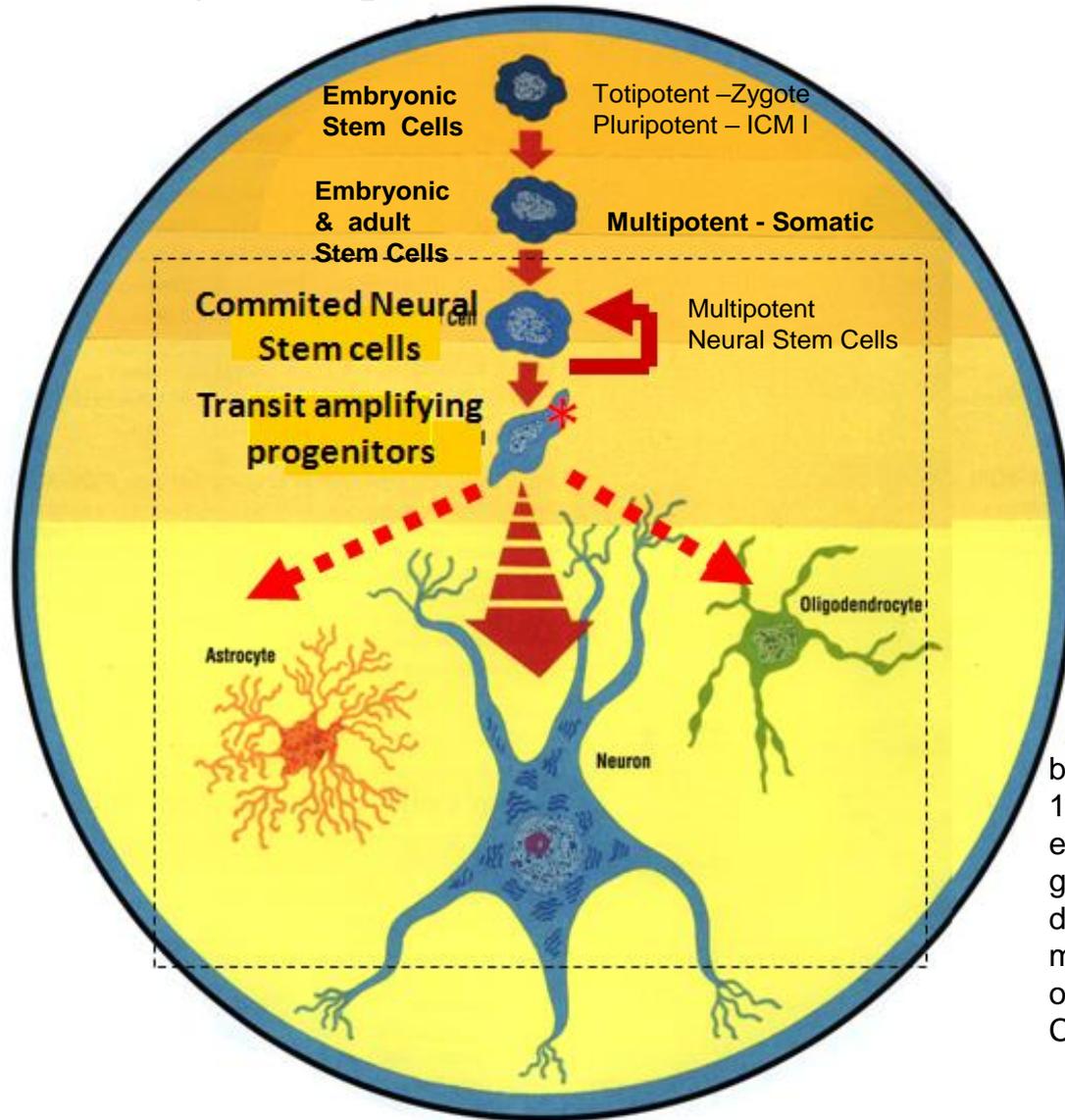
(A) Single-cell asymmetry



(B) Population asymmetry (symmetrical differentiation)



Unfolding Developmental Potential of Stem Cells



brain contains approximately 170 billion cells (neurons an equal number of glial cells of of great variety of types). This diversity begins with the multipotent neuroepithelial cells of the neural tube Neural Stem Cells.

Adult stem cells (typically multipotent):

create restricted array of cells in culture and have a finite number of generations for self-renewal (contributes to aging)

- hematopoietic stem cells that function to generate all the cells of the blood,
- Germinal stem Cells (testes, ovarian)
- brain
- epidermis,
- muscle,
- teeth,
- gut,
- lung,
- cornea,
- etc.,

Figure 5.4 To divide or not to divide: overview of stem cell regulatory mechanisms – Niche Factors

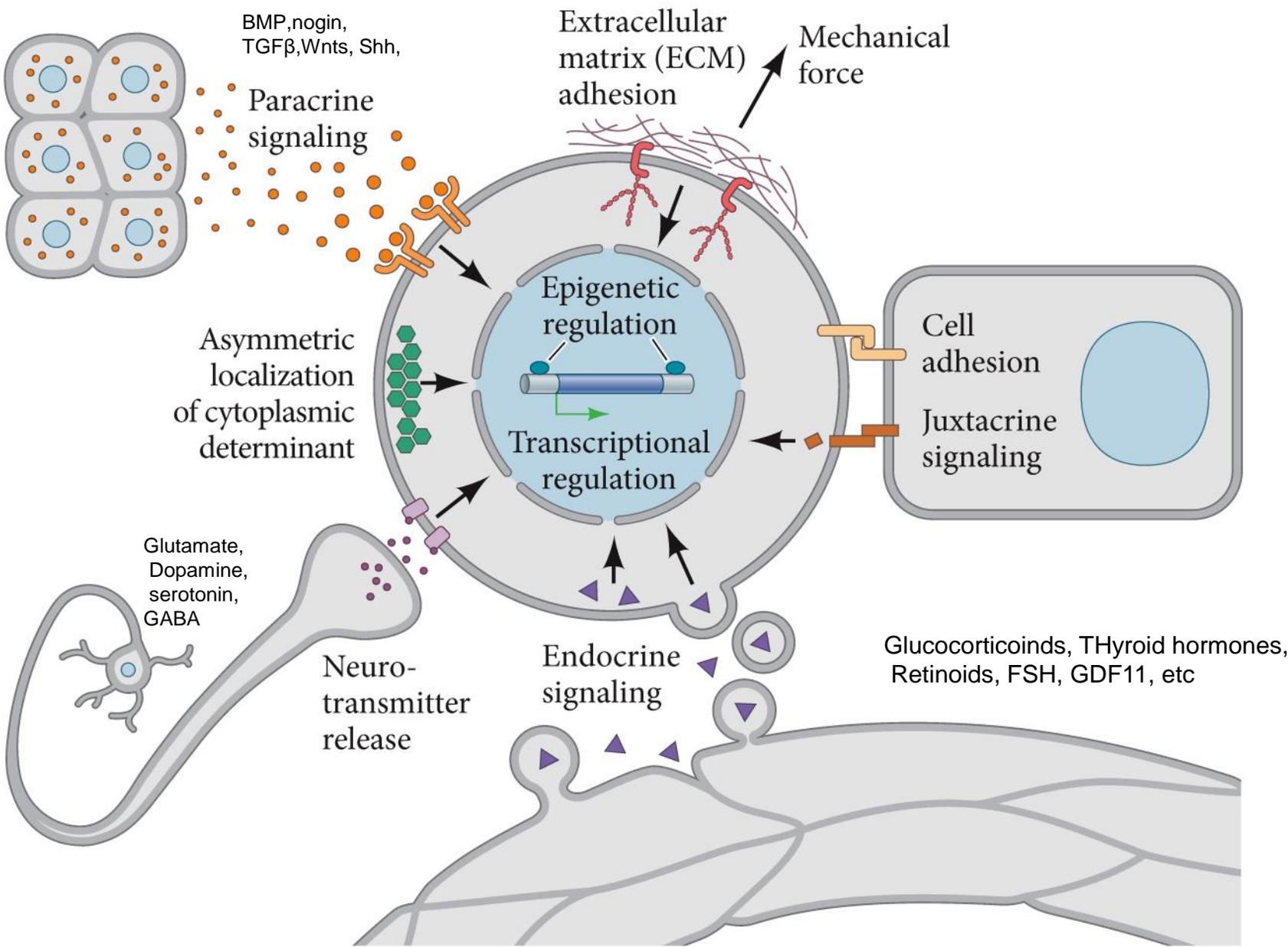


Figure 5.5 Establishment of the inner cell mass Niche in blastocyst

	Trophectoderm
	ICM
	Primitive endoderm
	Epiblast → Embryo

Embryonic placenta, chorion
Epiblast, hypoblast
Hypoblast > primitive endoderm, yolk sac
Epiblast > 200 cell types

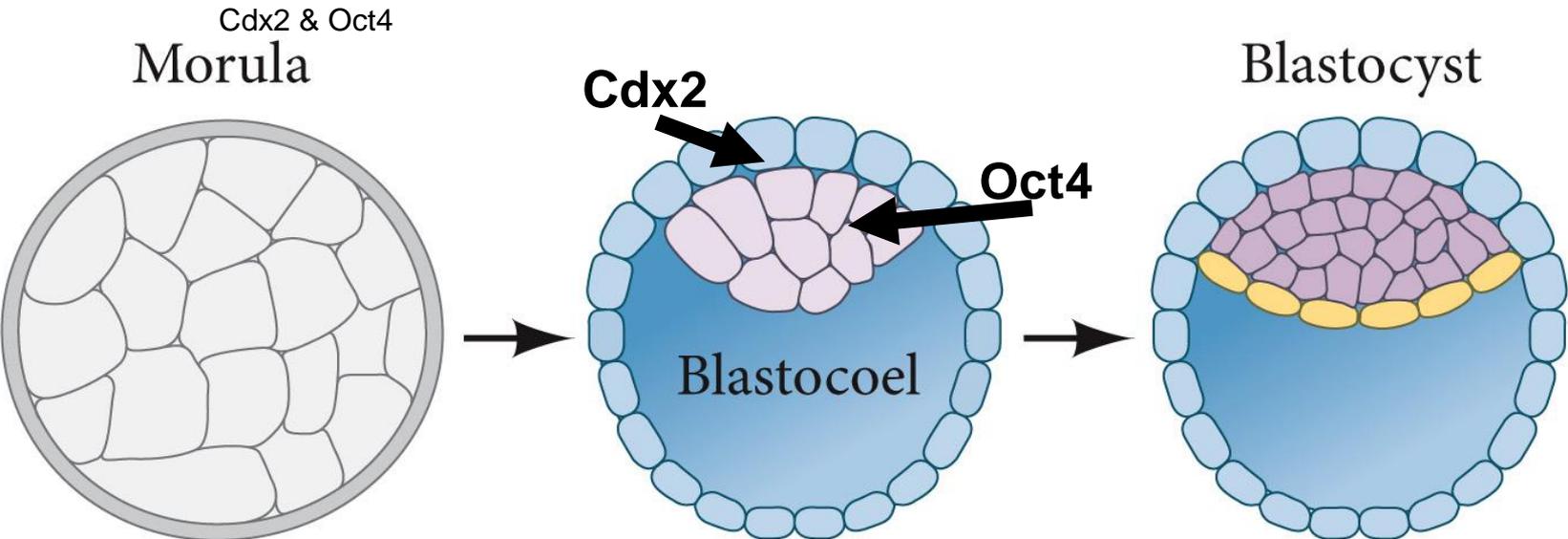


Figure 5.6 Divisions about the apicobasal axis

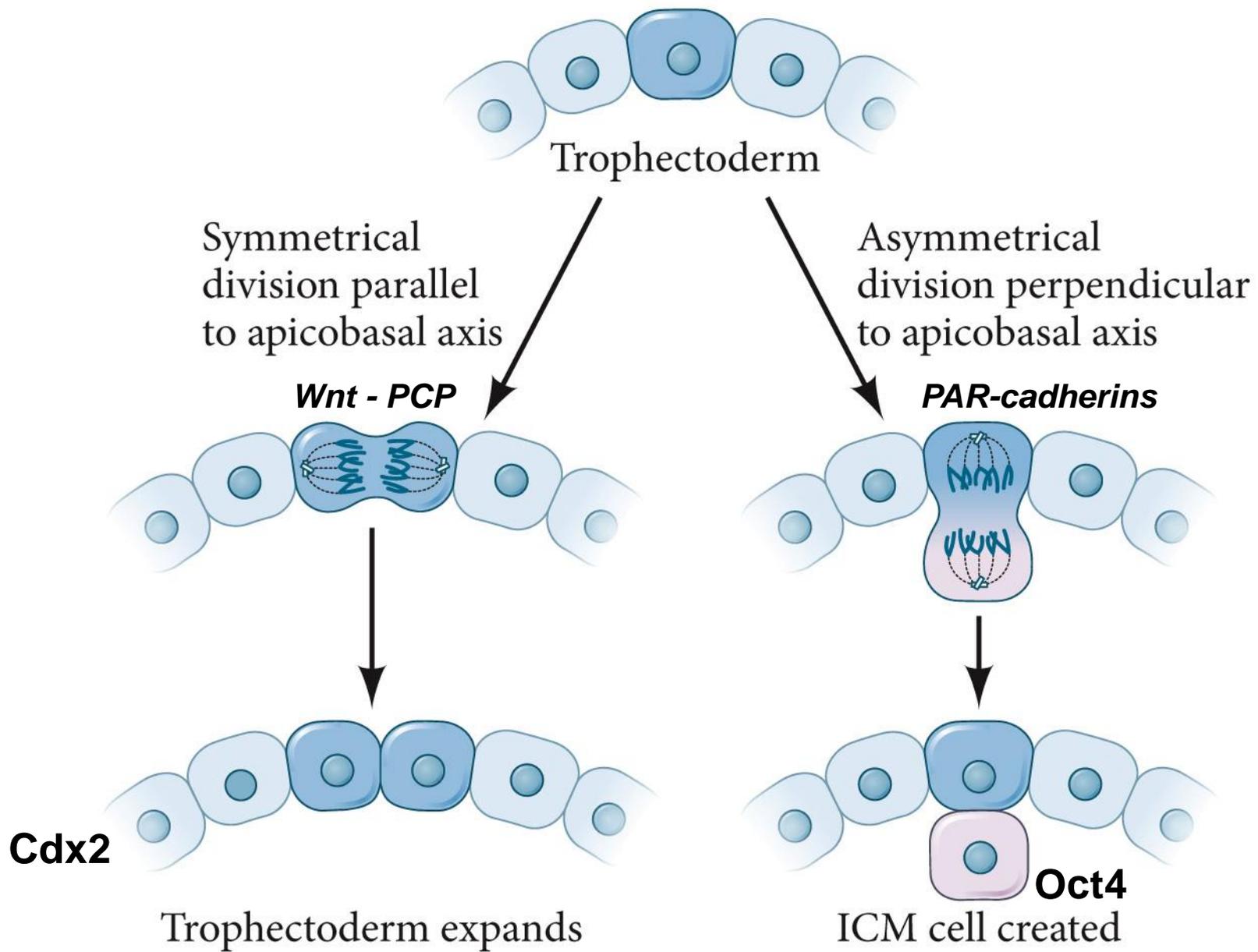
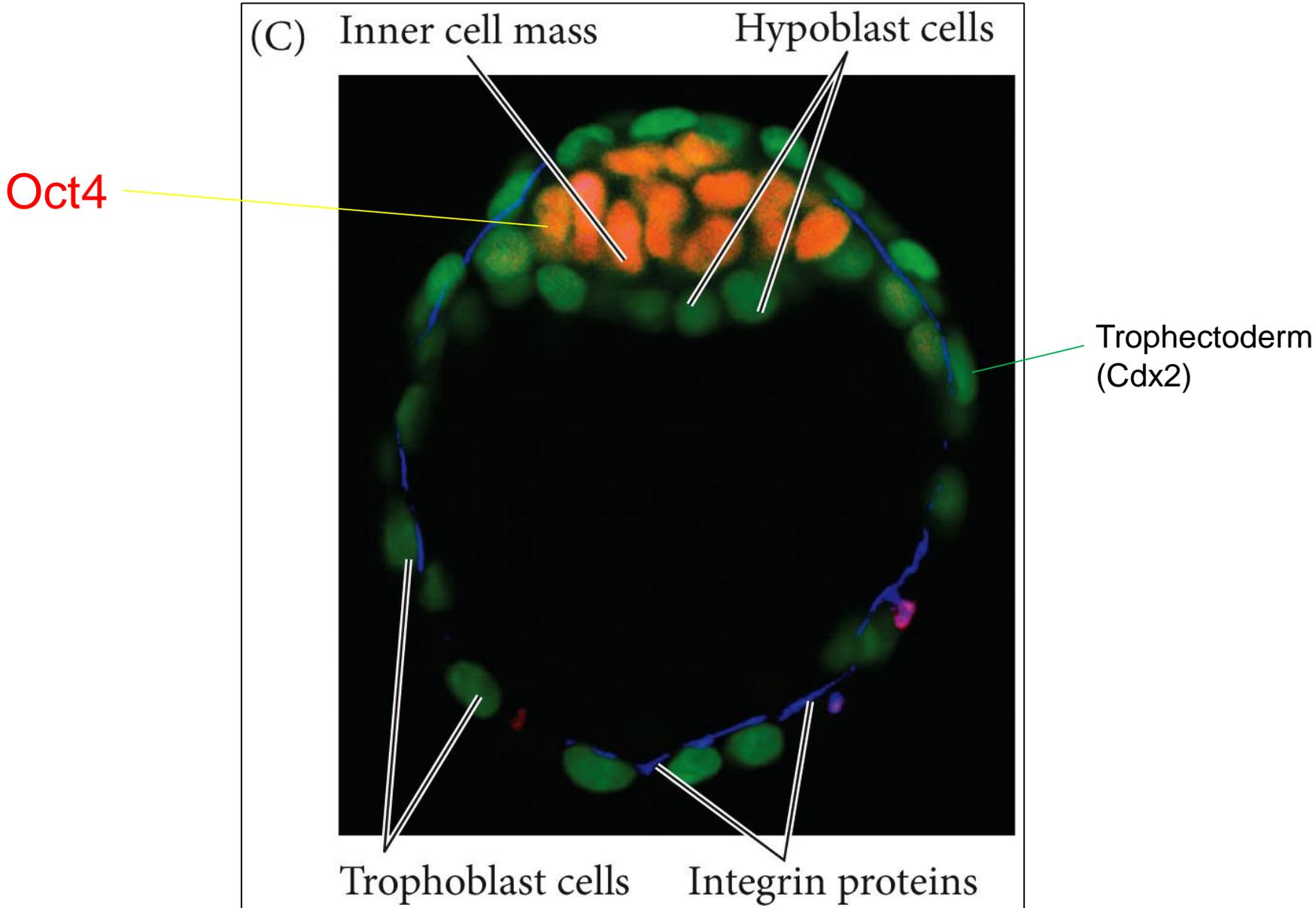


Figure 12.15 Possible pathway initiating the distinction between inner cell mass and trophoblast (Part 3)



Apicobasal partitioning in Morula

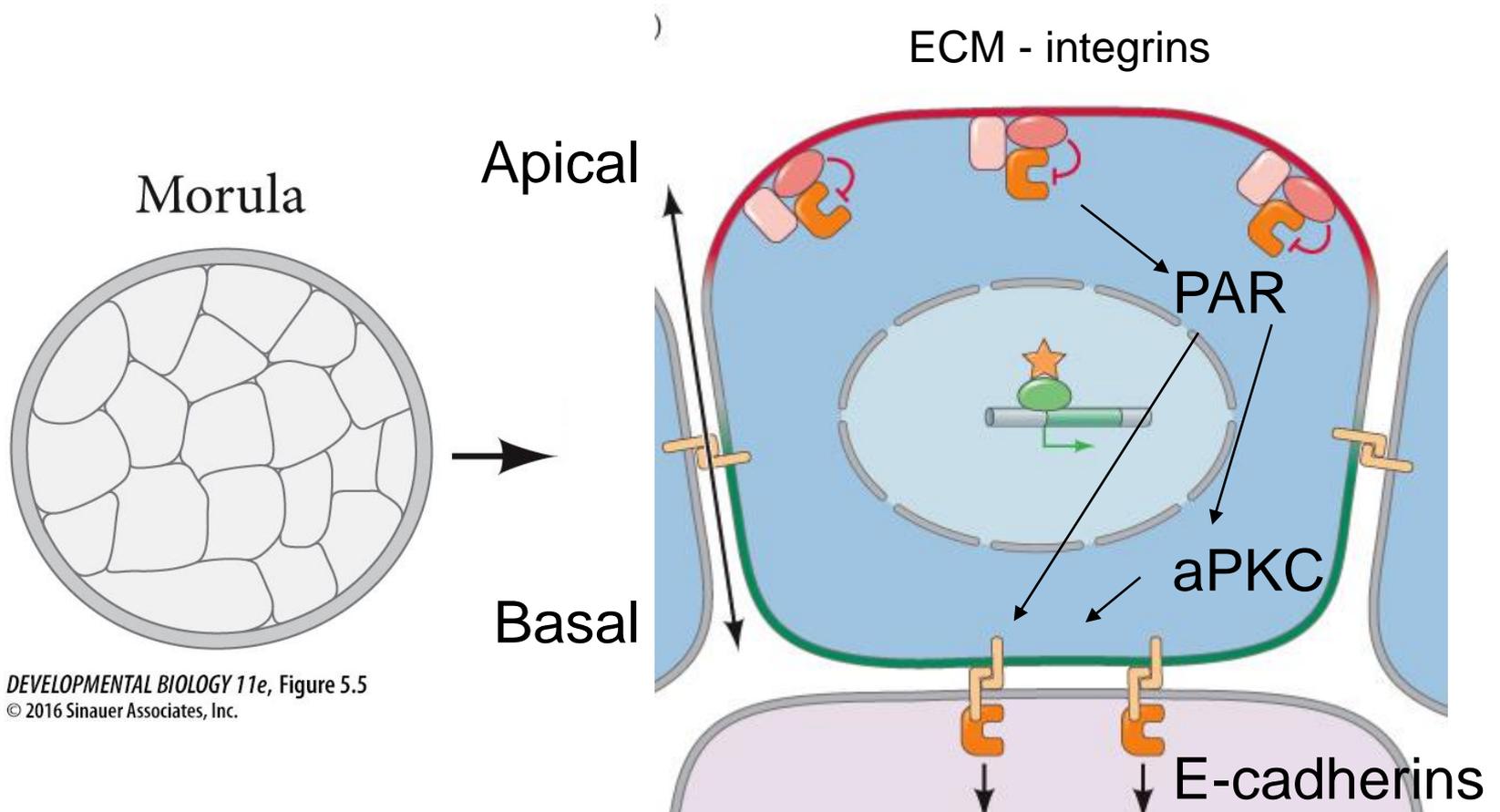
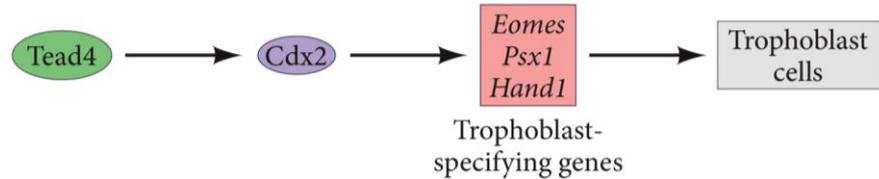
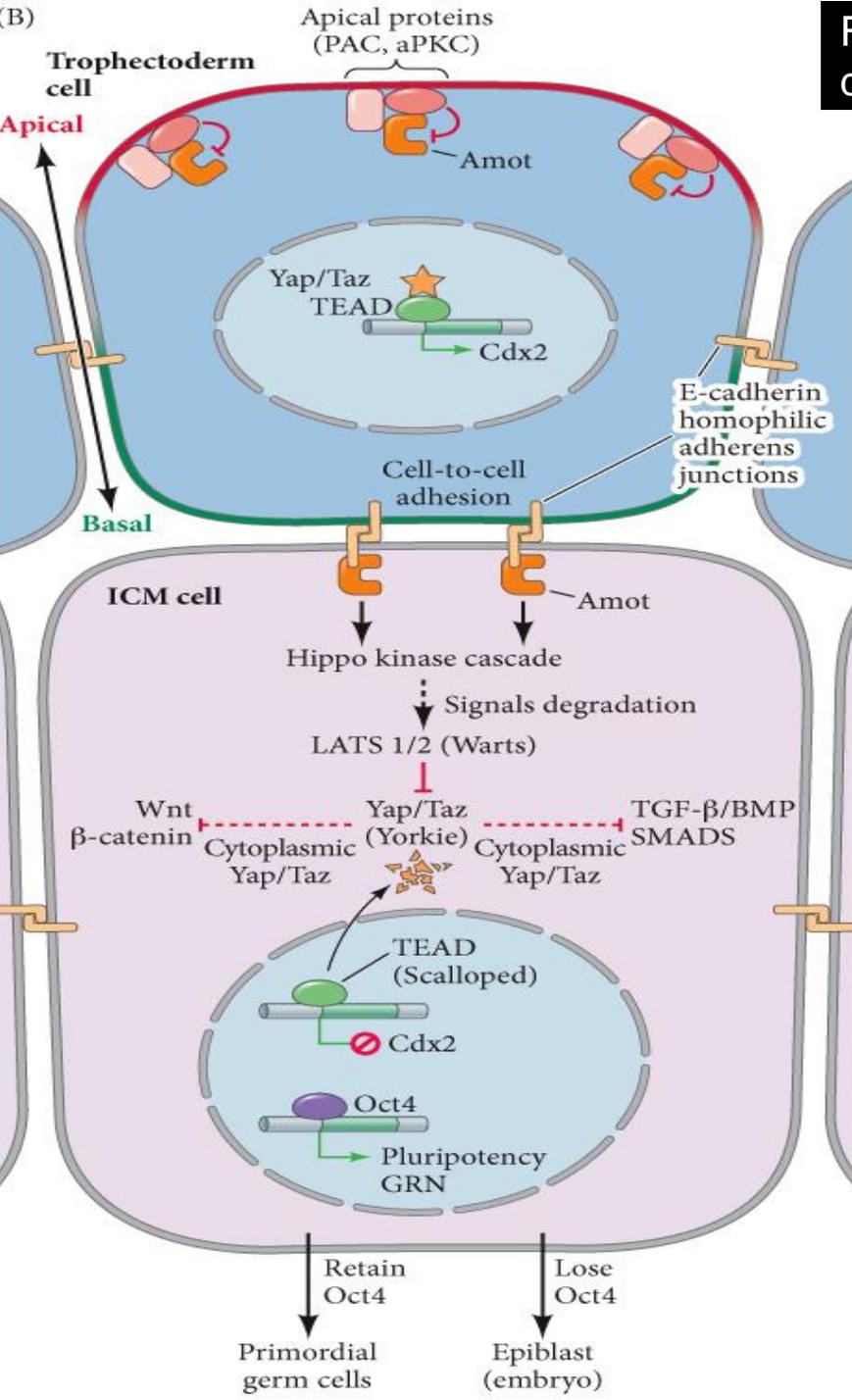
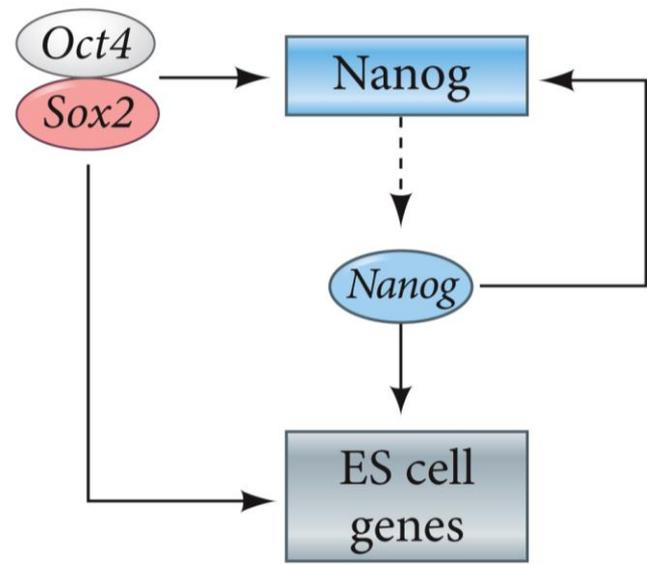


Figure 5.7 Hippo - Possible pathway initiating the distinction between inner cell mass and trophoblast

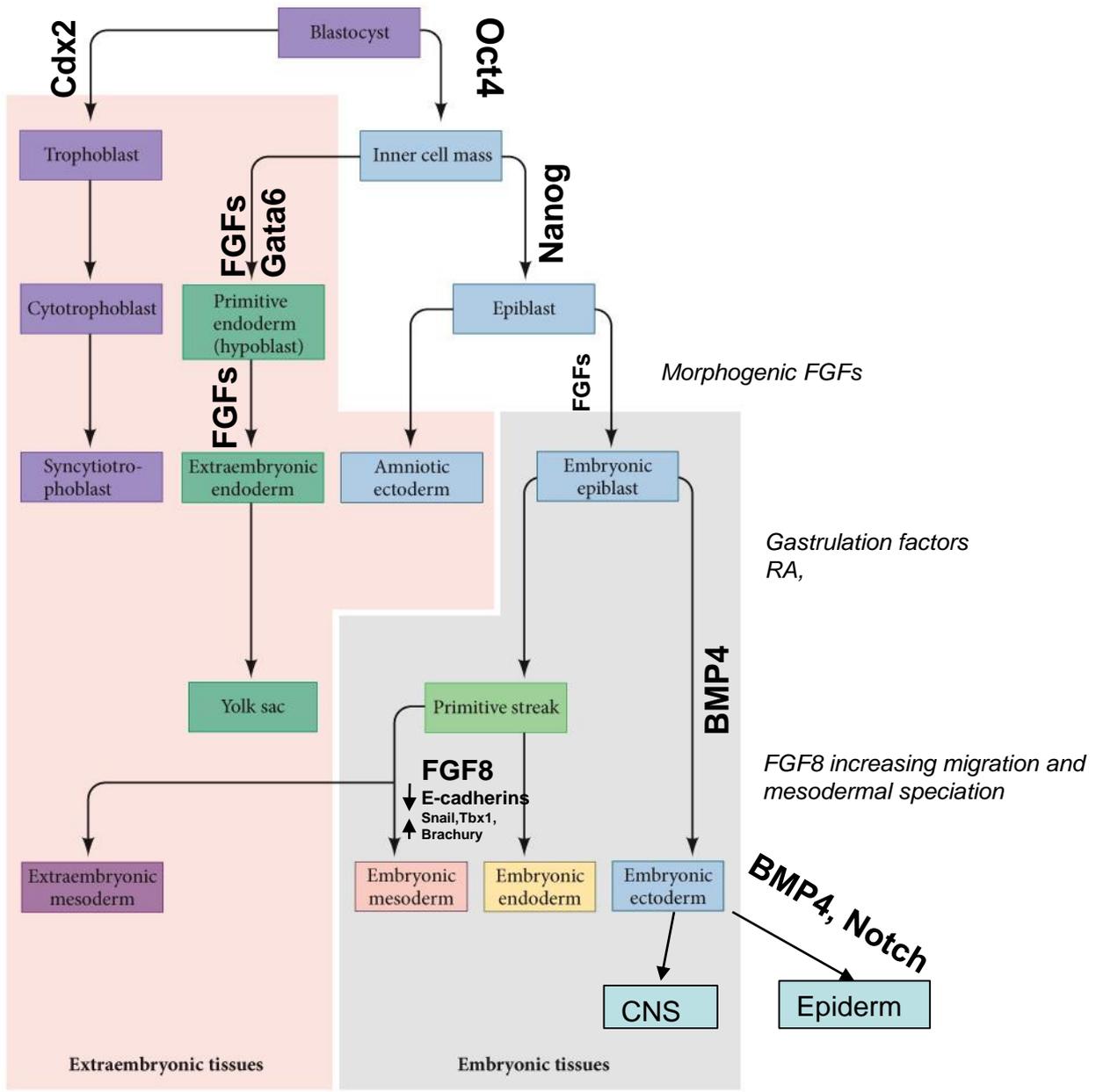


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DEVELOPMENTAL BIOLOGY 11e, Figure 12.14 (Part 1)
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Figure 12.17 Tissue and germ layer formation in the early human embryo (Part 2)



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Table 5.1 Some stem cell niches of adult humans

TABLE 5.1 Some stem cell niches of adult humans

Stem cell type	Niche location	Cellular components of niche
LOW TURNOVER^a		
Brain (neurons and glia)	Ventricular-subventricular zone (V-SVZ; see Figure 5.10), subgranular zone	Ependymal cells, blood vessel epithelium
Skeletal muscle	Between basal lamina and muscle fibers	Muscle fiber cells
HIGH TURNOVER^a		
Mesenchymal stem cells (MSCs)	Bone marrow, adipose tissue, heart, placenta, umbilical cord	Probably blood vessel epithelium
Intestine	Base of small intestinal crypts (see Figure 5.13)	Paneth cells, MSCs
Hematopoietic (blood-forming) stem cells (HSCs)	Bone marrow (see Figure 5.15)	Macrophages, T _{reg} cells, osteoblasts, pericytes, glia, neurons, MSCs
Epidermis (skin)	Basal layer of epidermis	Dermal fibroblasts
Hair follicle	Bulge (see Figure 16.17)	Dermal papillae, adipocyte precursors, subcutaneous fat, keratin
Sperm	Testes	Sertoli cells (see Figure 6.21)

^a Niches with low rates of cell turnover generate stem cells for repair, slow growth, and (in the case of neurons) learning. Niches with high turnover are constantly producing new cells for bodily maintenance.

Figure 5.8 Stem cell niche in *Drosophila* testes (Part 1)

(A)

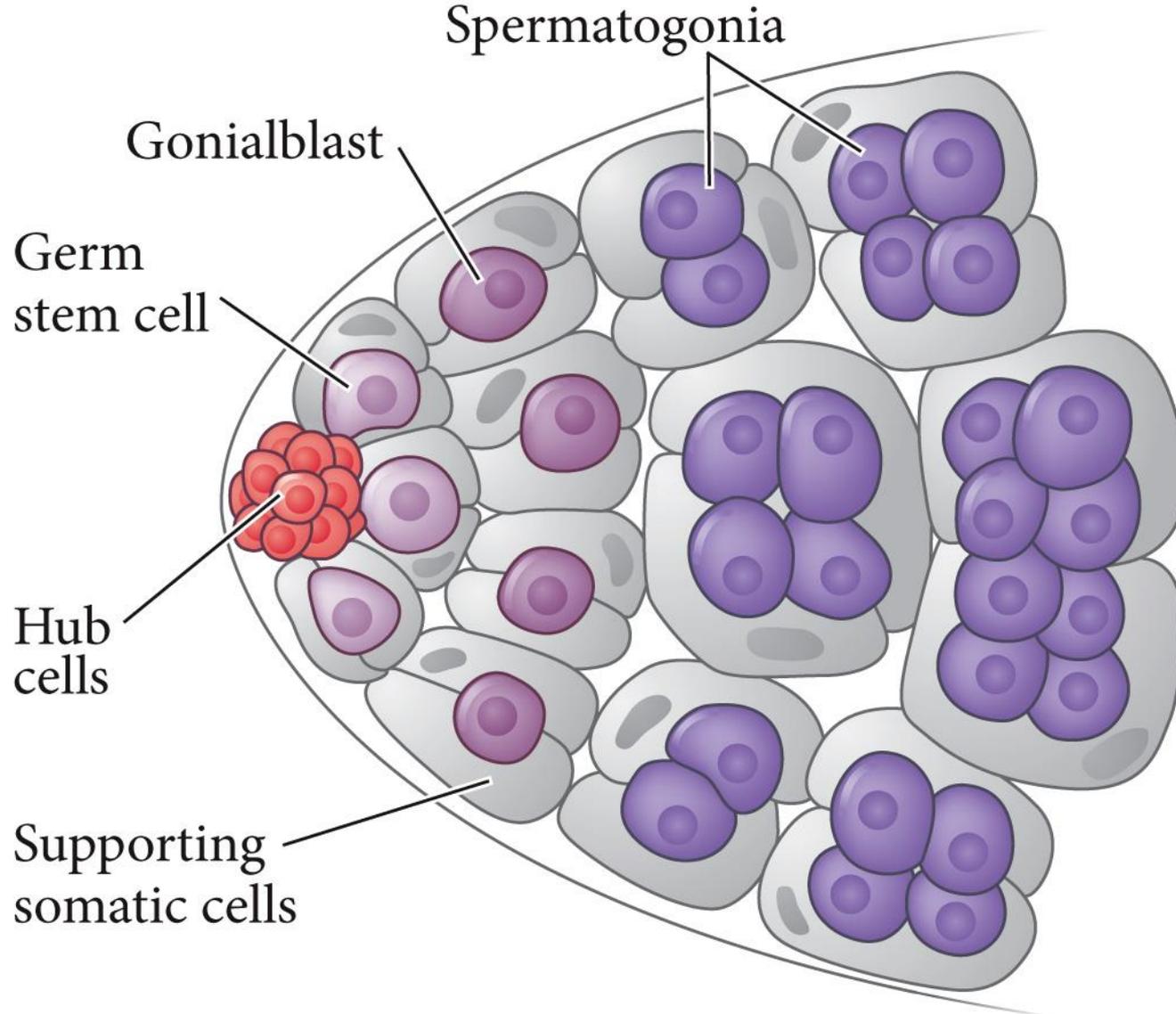
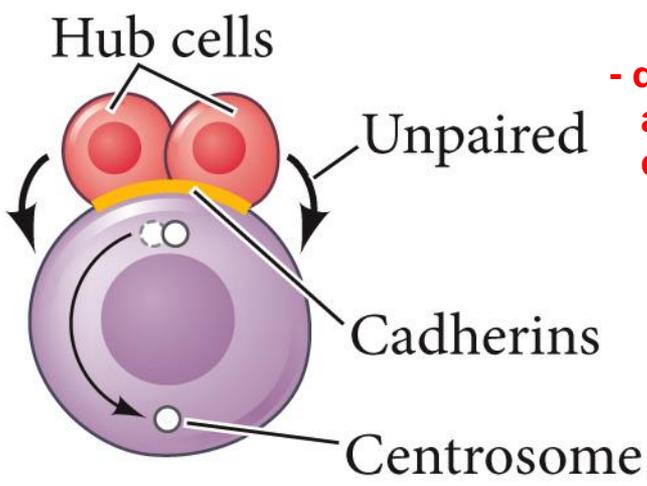
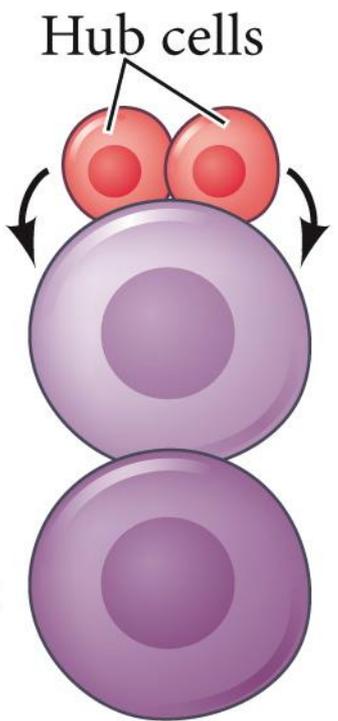


Figure 5.8 Hub cells "Unpaired" dictates an assymetrical fate of the Stem cell progeny

(C)



- dictates centrosome migration and mitotic plane, cadherin distribution



Unpaired proximal cell effect >JAK-STAT>pluripotency, >restores Cadherin

Committed goniblast > spermatozoid

Unpaired gene expressed in
Hub cells
Reporter β -galactosidase
inserted into the gene for
Unpaired reveals that this
protein is transcribed in the
somatic hub cells.

(B)

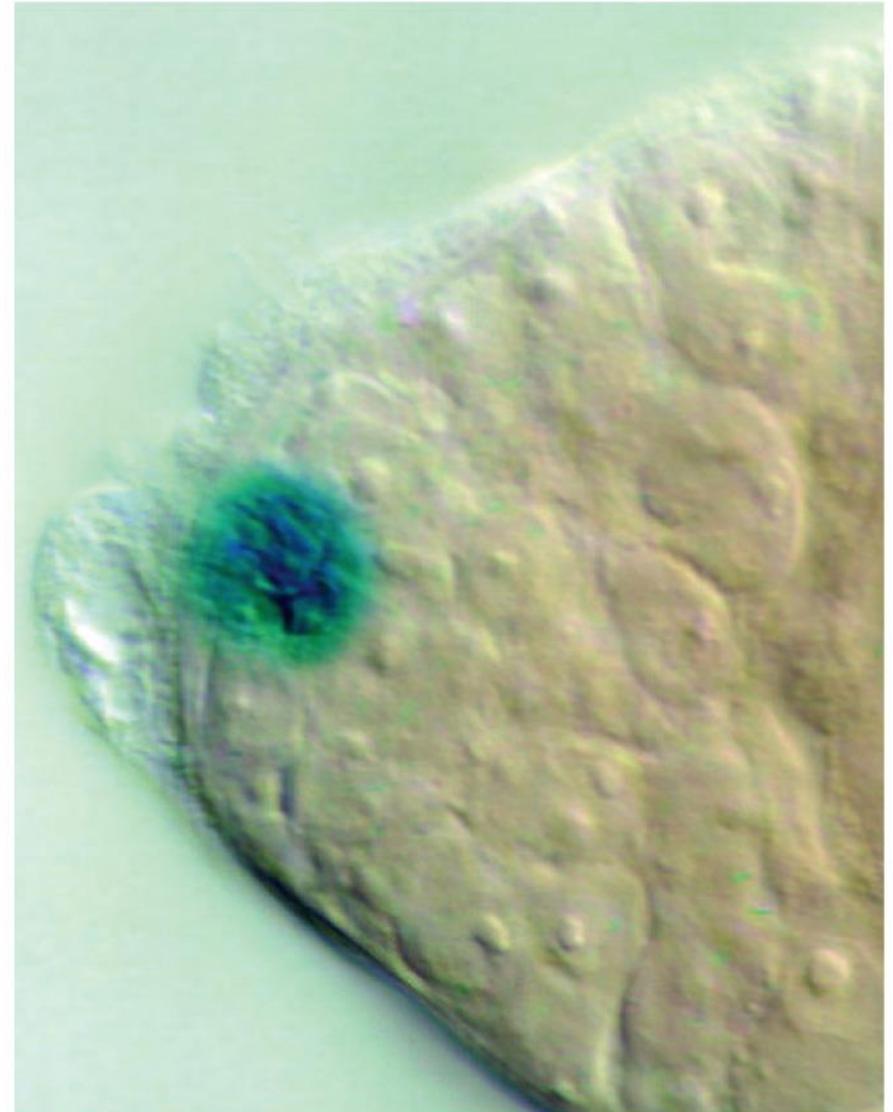
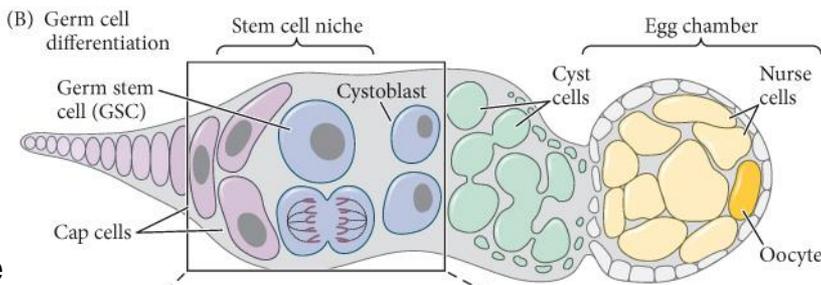
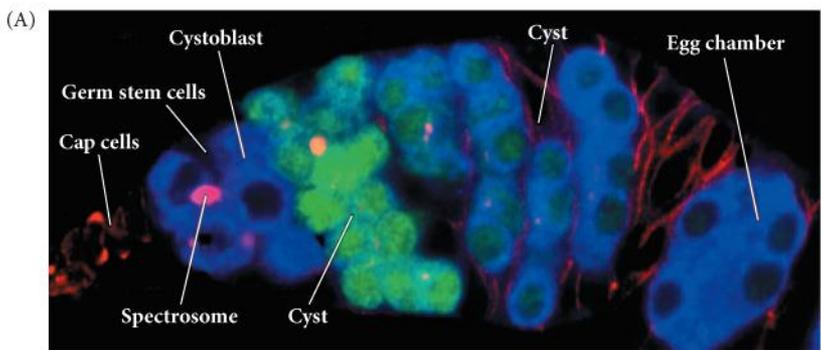


Figure 5.9 *Drosophila* ovarian stem cell niche germanium.



E-cadh arrests GSC in space
BMP prevents differentiation

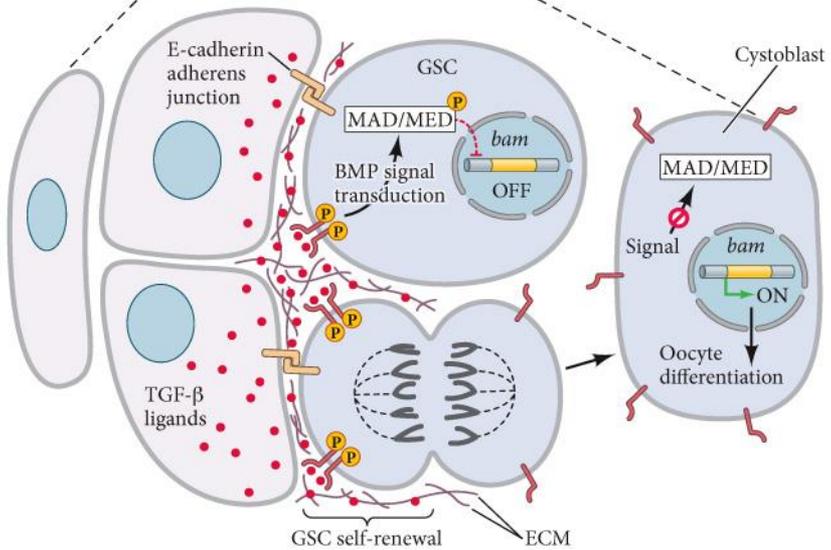
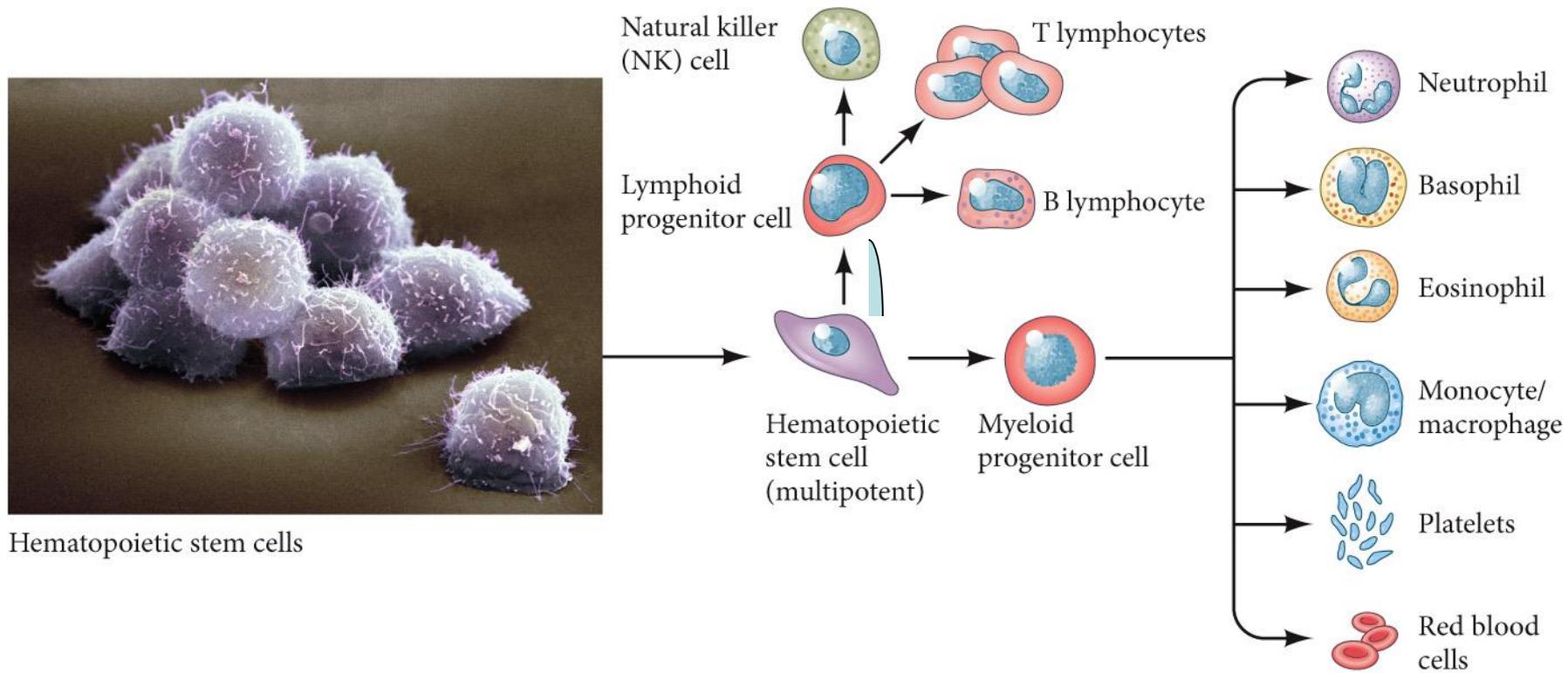


Figure 5.3 Blood-forming (hematopoietic) stem cells (HSCs)

Best know adult stem cells - Hematopoietic multipotent stem cells > all blood cells

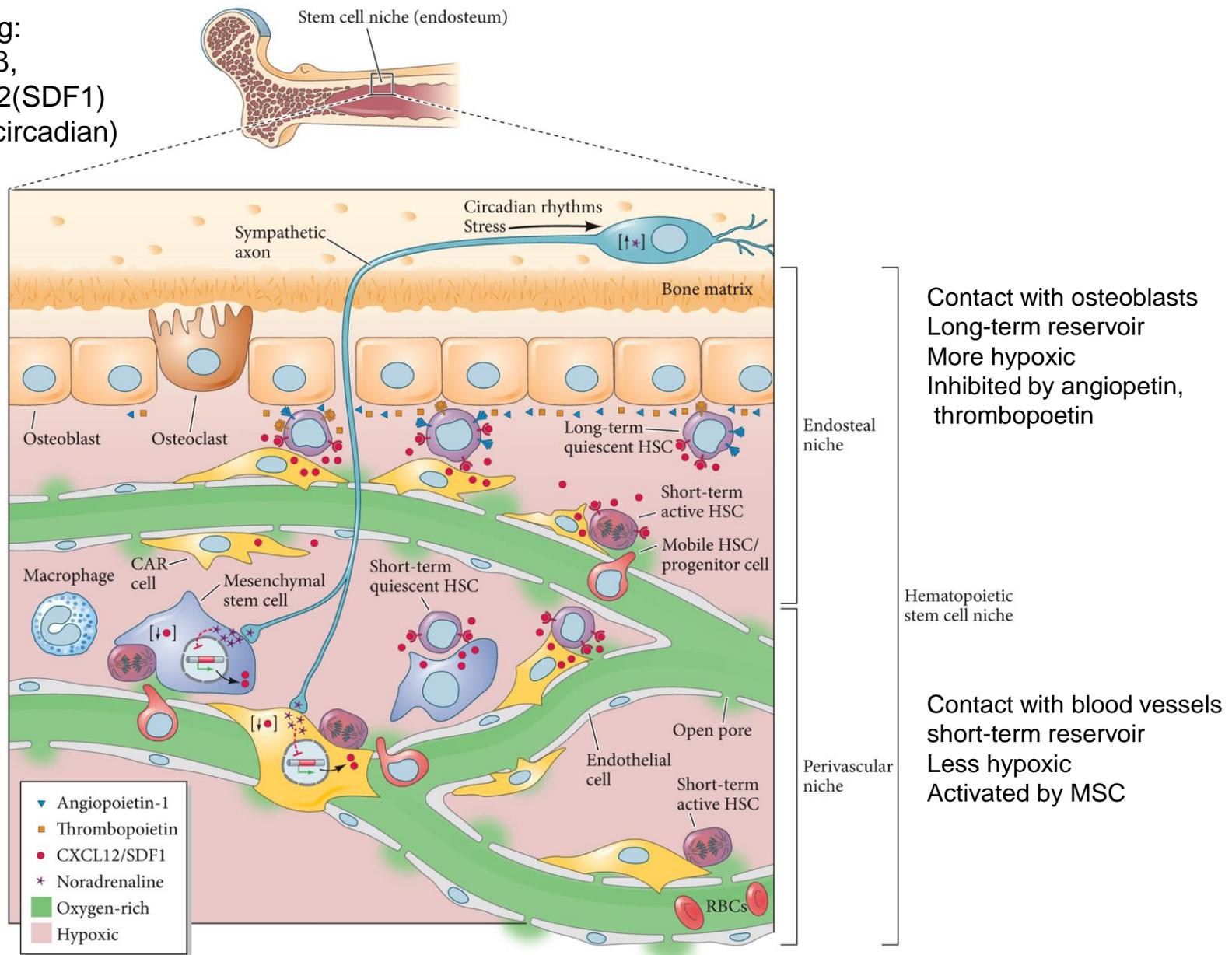


Hematopoietic stem cells

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Figure 5.15 Model of adult HSC Hematopoietic Stem Cell niche

Factors controlling:
 Wnt, Notch, TGFβ,
 integrins, CXCL12(SDF1)
 Norepinephrine (circadian)



Mesenchymal Stem Cell : Supporting a variety of adult Tissues

Multiple Sources and Niches:

- Bone marrow (original finding - bone marrow derived stem cells)
- dermis of the skin, bone, fat, cartilage, tendon, muscle, thymus, cornea, and dental pulp, umbilical cord and placenta
- MSC - “Split personality” as (1) supportive stromal cells secreting ECM and (2) self-renewing stem cells on the other.

Clonal plasticity

- In culture MSC clonal populations can form different organs, examples: osteoblasts (green) and adipocytes (red)
- **Paracrine control:**
 - PDGF & TGF- β signaling > chondrogenesis,
 - FGF > bone cells

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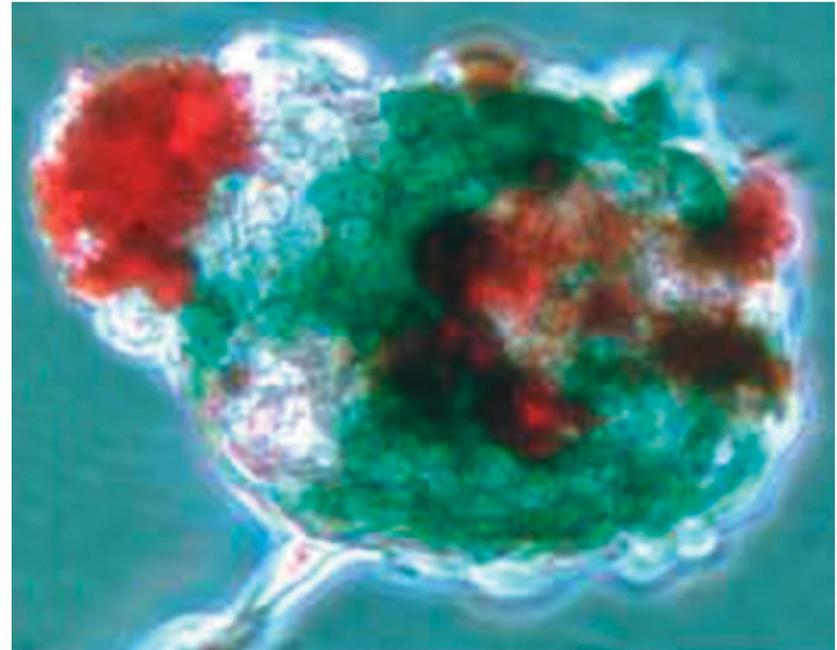
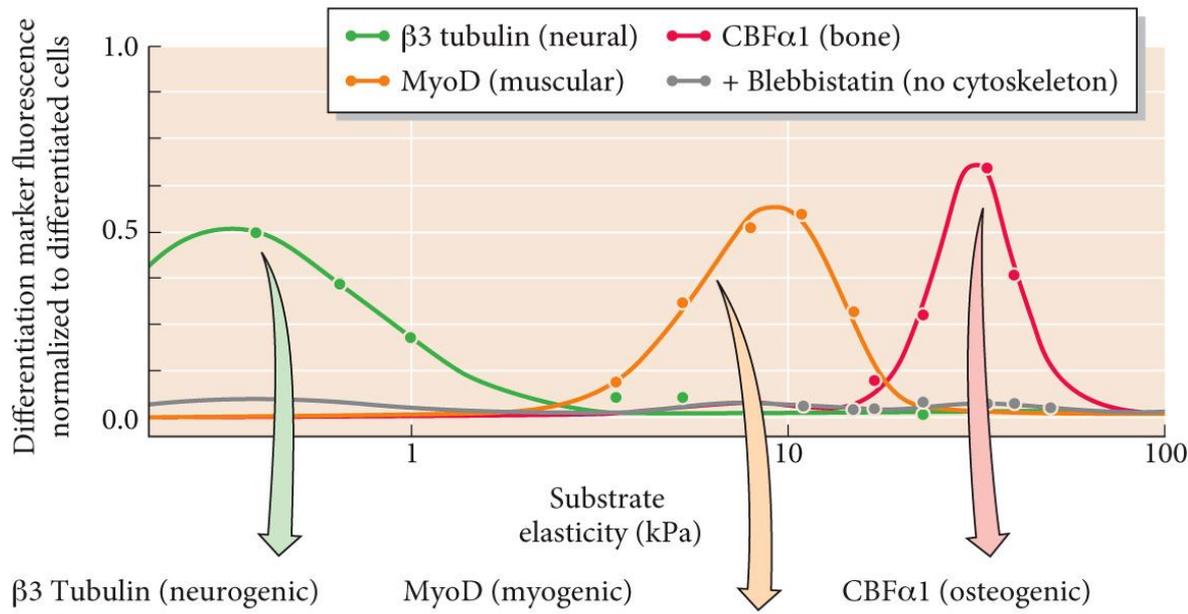


Figure 5.17 A mesensphere containing two derived cell types

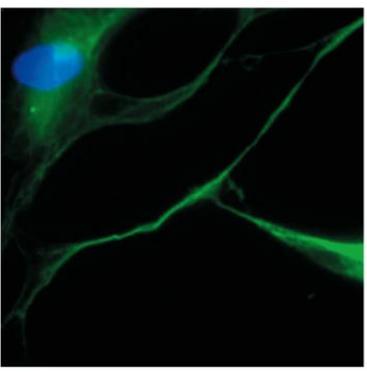
Figure 5.18 Mesenchymal stem cell differentiation is influenced by the elasticity of the matrices upon which the cells sit

ECM laminin keep MSCs in a state of undifferentiated "stemness"
Physical matrix elasticity controls cell differentiation

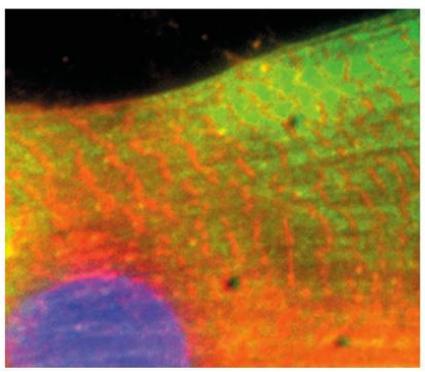


collagen-coated gels of different elasticity (transduced via microfilament – blocked by blebbistatin)

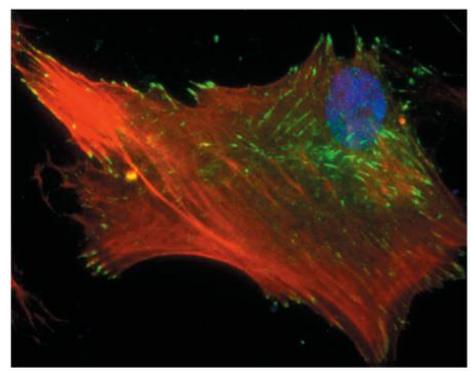
$\beta 3$ Tubulin (neurogenic)



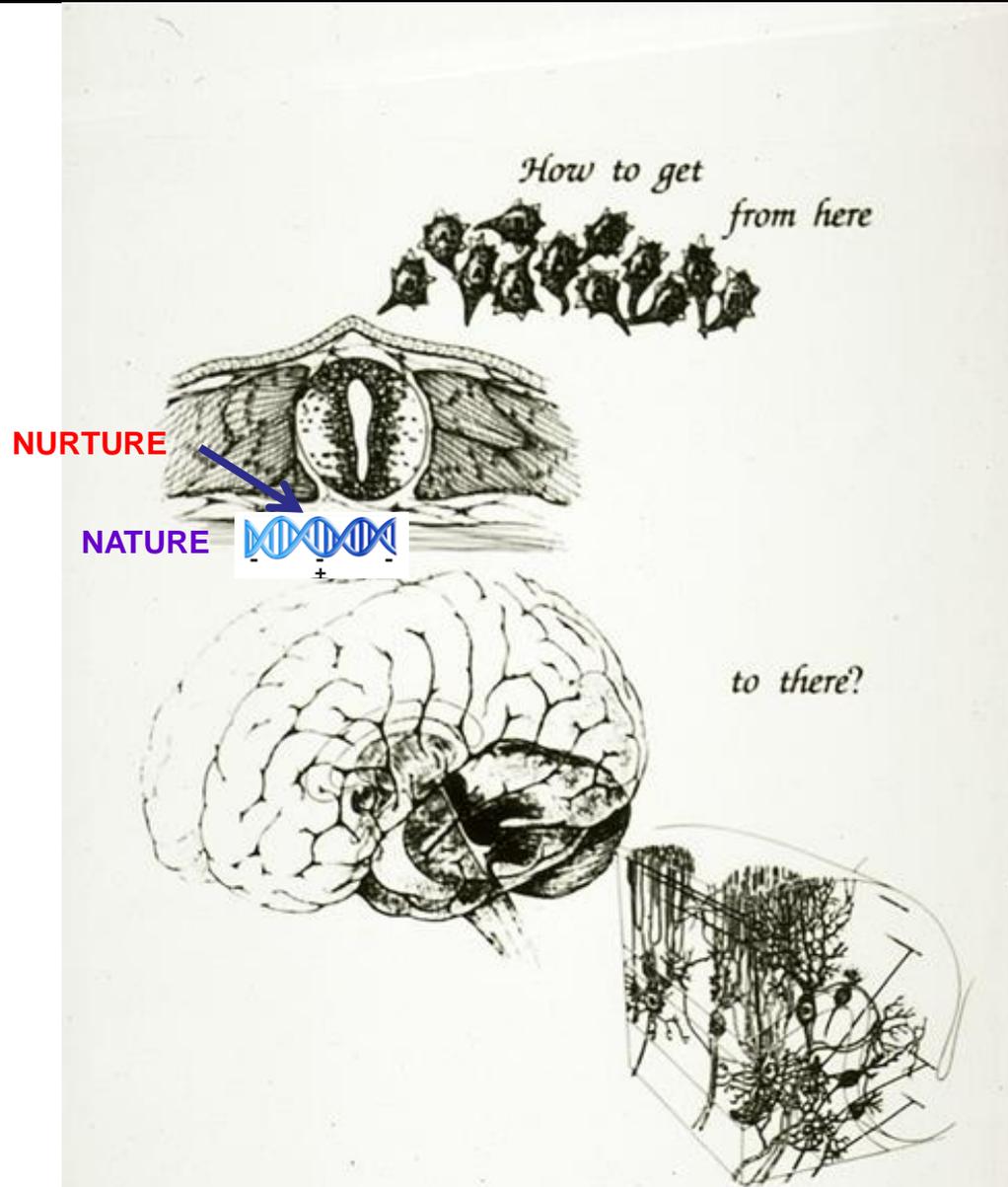
MyoD (myogenic)



CBF $\alpha 1$ (osteogenic)



Neural Stem Cells Niche in Developing CNS



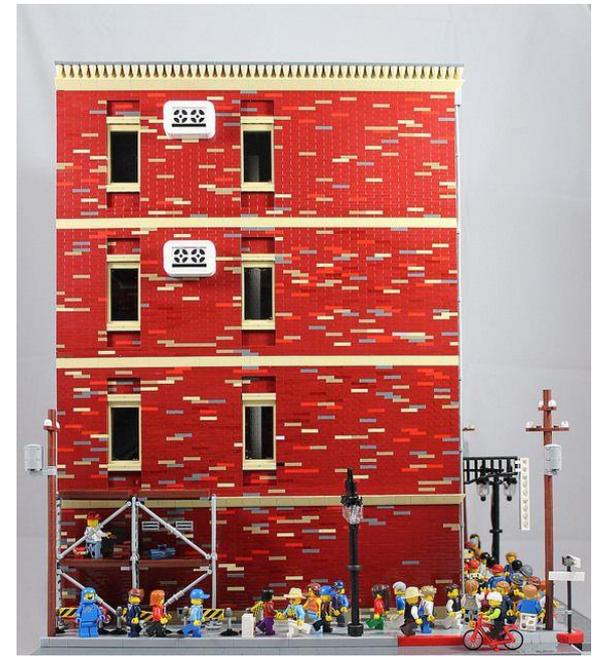
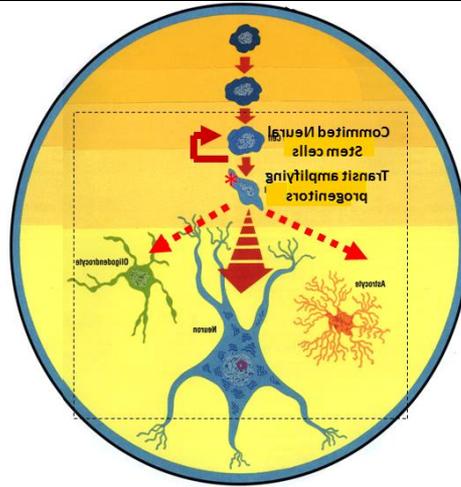
Can cells proliferate to make new neurons in adult brain?

NO - new neurons be made Santiago Ramon y Cajal's dogma: "Everything may die, nothing may be regenerated"

YES - (Altman and Das 1965; Nottebohm 1985) reported the occurrence of adult neurogenesis in rats, cats, and birds' brains as early as 1962.

NO - (Rakic 1985; Eckenhoff and Rakic 1988) - had tried to identify, to no avail, stem cells in the adult brains of higher primates

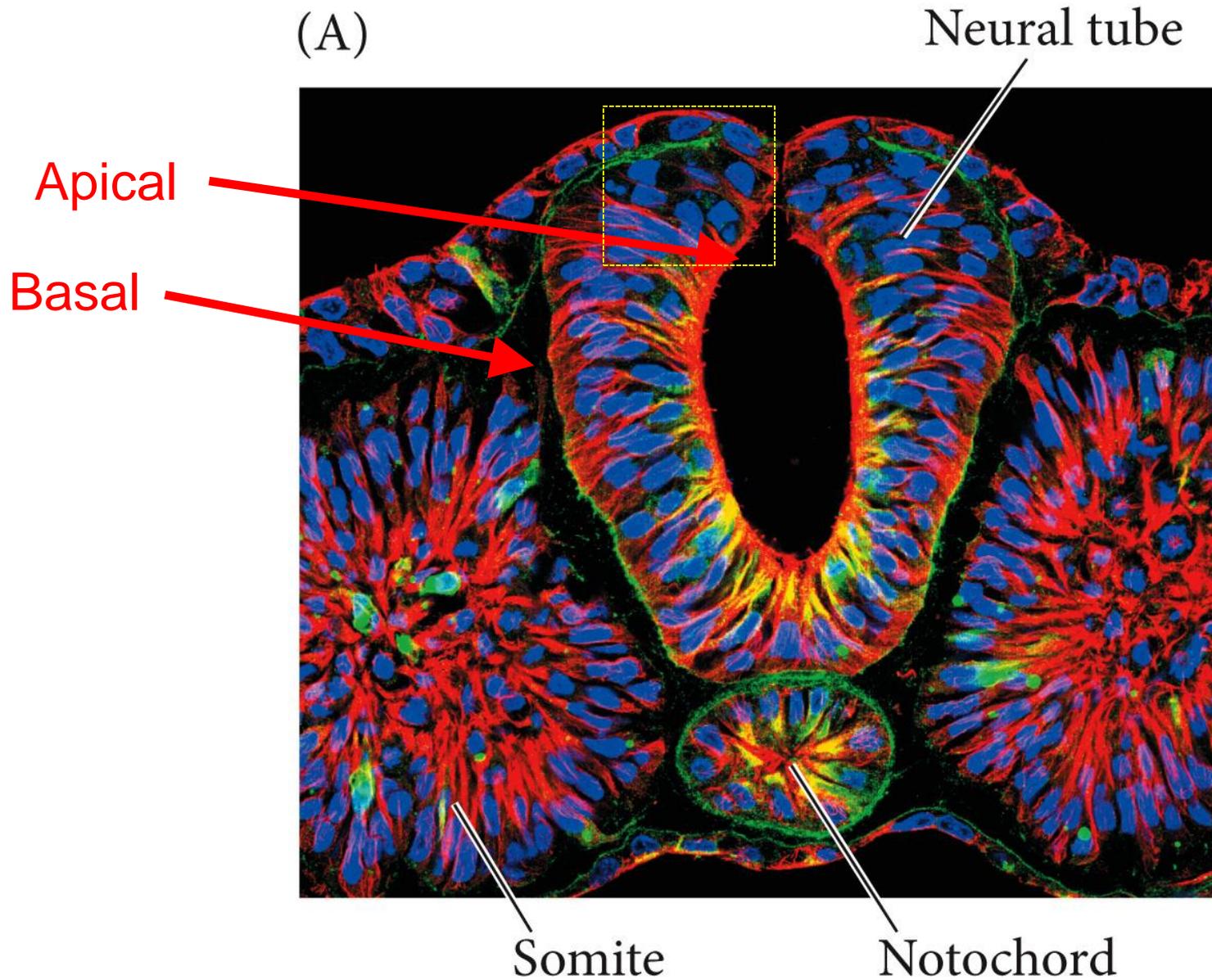
YES - (Eriksson et al. 1998) discovered an in vivo neurogenesis occurring in the adult human brain by injecting a non-radioactive Bromodeoxy Uridine (BrdU), a synthetic analog of thymidine, to monitor neuronal proliferation in terminal cancer patients. BrdU labelled cells expressed neuronal markers



NCS: Neuroepithelial cells
ventricular
Radial Glial

subventricular (outer)
Radial Glia

Figure 1.9 Two types of microscopy are used to visualize the notochord and its separation of vertebrate embryos into right and left halves (Part 1)



Neuroepithelial (stem) cells (NEC):

- the first multipotent neural stem cells of the embryo, make up the neural plate and early neural tube (later transform to radial glia cells RGC),
- NEC are polarized along their apical to basal axis, single spans the tube wall.
- the apical NEC surface borders the internal cavity to be filled with the CSF in neural tube,
- NEC basal surface forms an endfoot - swelling of its basal membrane, pial matter a fibrous membranes that surround nervous tissues.

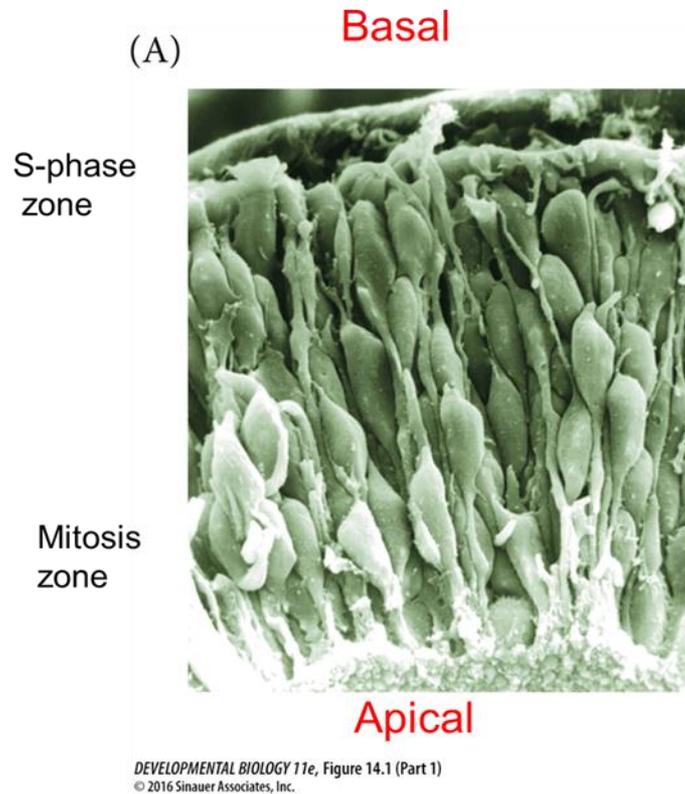
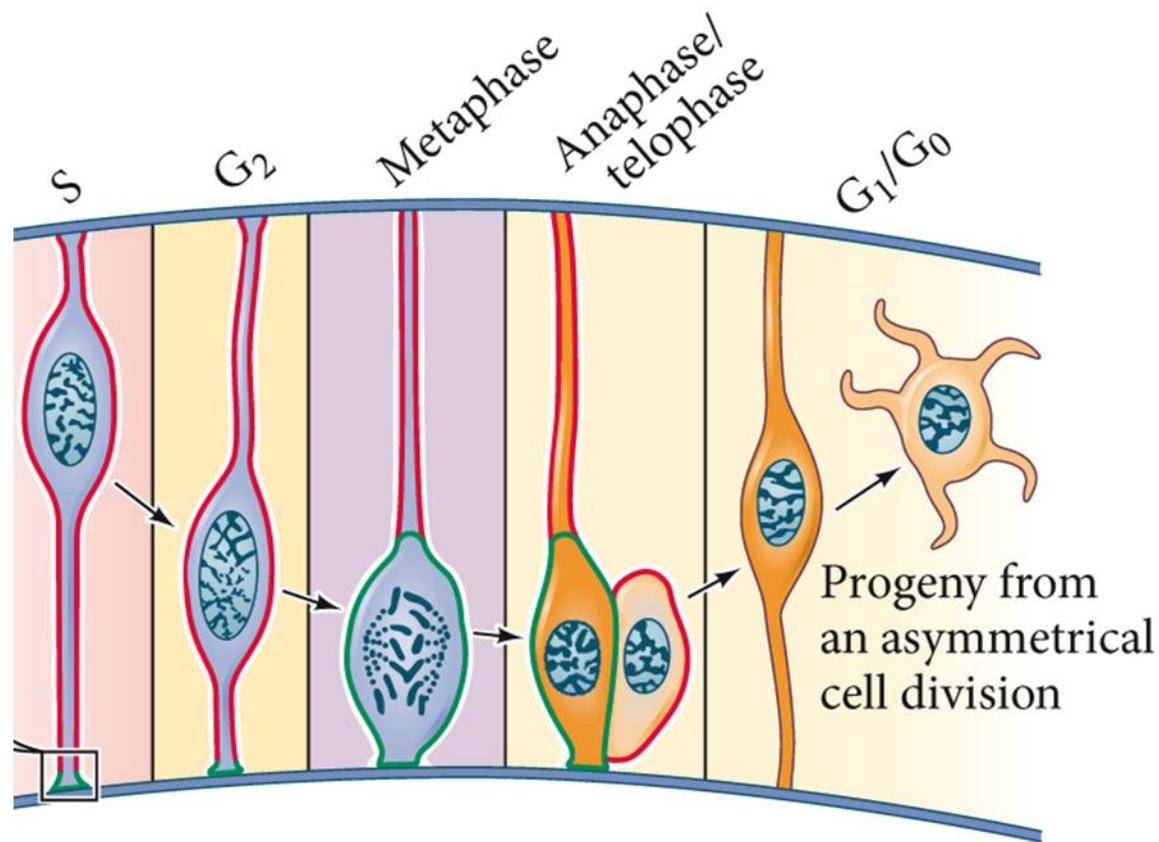
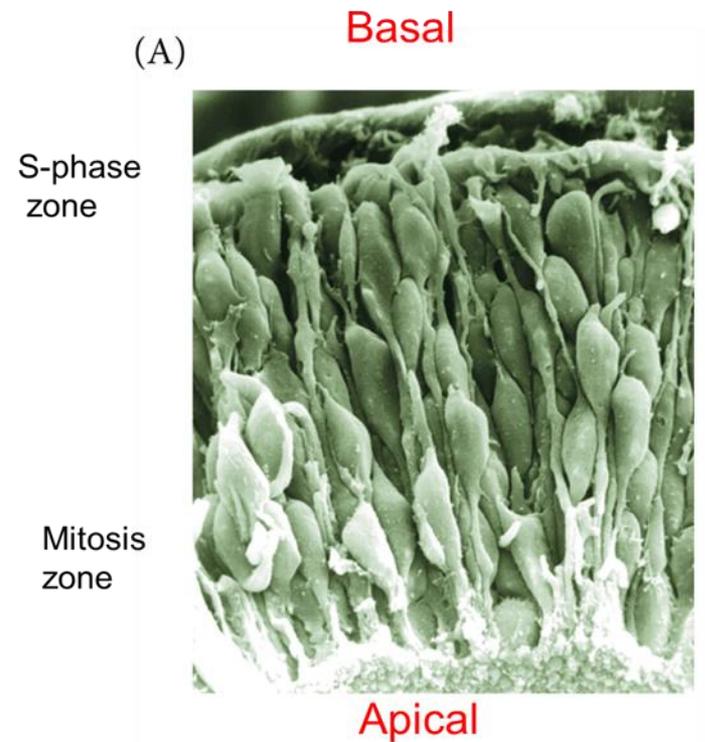


Figure 14.1 Cell types of the CNS. (A) Scanning electron micrograph of a newly formed chick neural tube, showing neuroepithelial cells at different stages of their cell cycles spanning the full width of the epithelium.

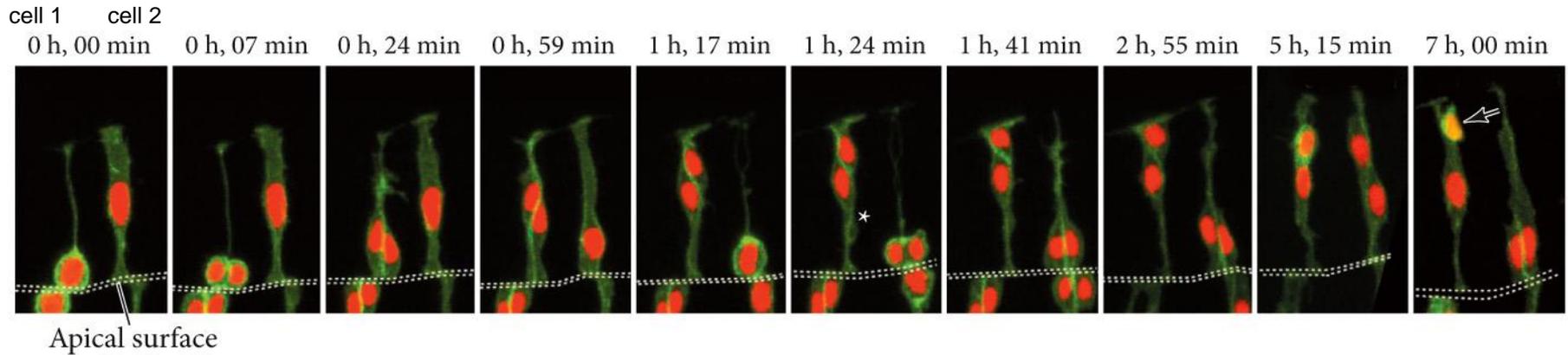
Microtubule and dynein dependent interkinetic nuclear migration (Fig. 14.1A; 14.1)



Asymmetric (shown) or symmetric division

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asymmetrical (1) and symmetrical (2) division



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Figure 14.9 Live imaging of neuroepithelial cell interkinetic nuclear migration and cell division cells zebrafish embryonic hindbrain. Two adjacent progenitor cells in the germinal epithelium were recorded over 7 hours (cell membranes (green) and nuclei (red)). A reporter gene specifically marks neurons (yellow).

Cell 1 – asymetric division,
Cell 2 – symmetric division

Figure 14.14 symmetrical versus asymmetrical division depends on the **plane of division**
Mitotic spindle parallel – **symmetric**
Mitotic spindle oblique - **asymmetric**

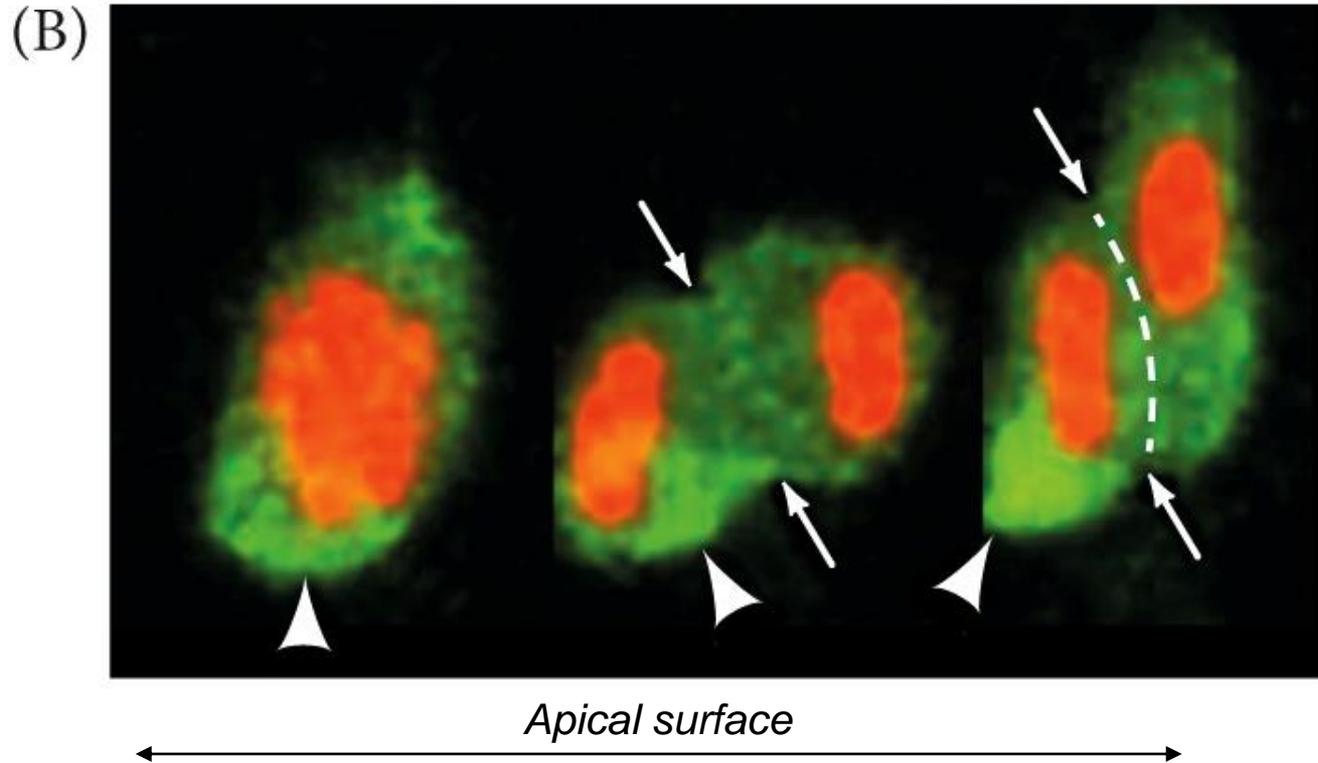


Figure 14.14 Asymmetrical division of radial glia mediated by Par3 and Notch

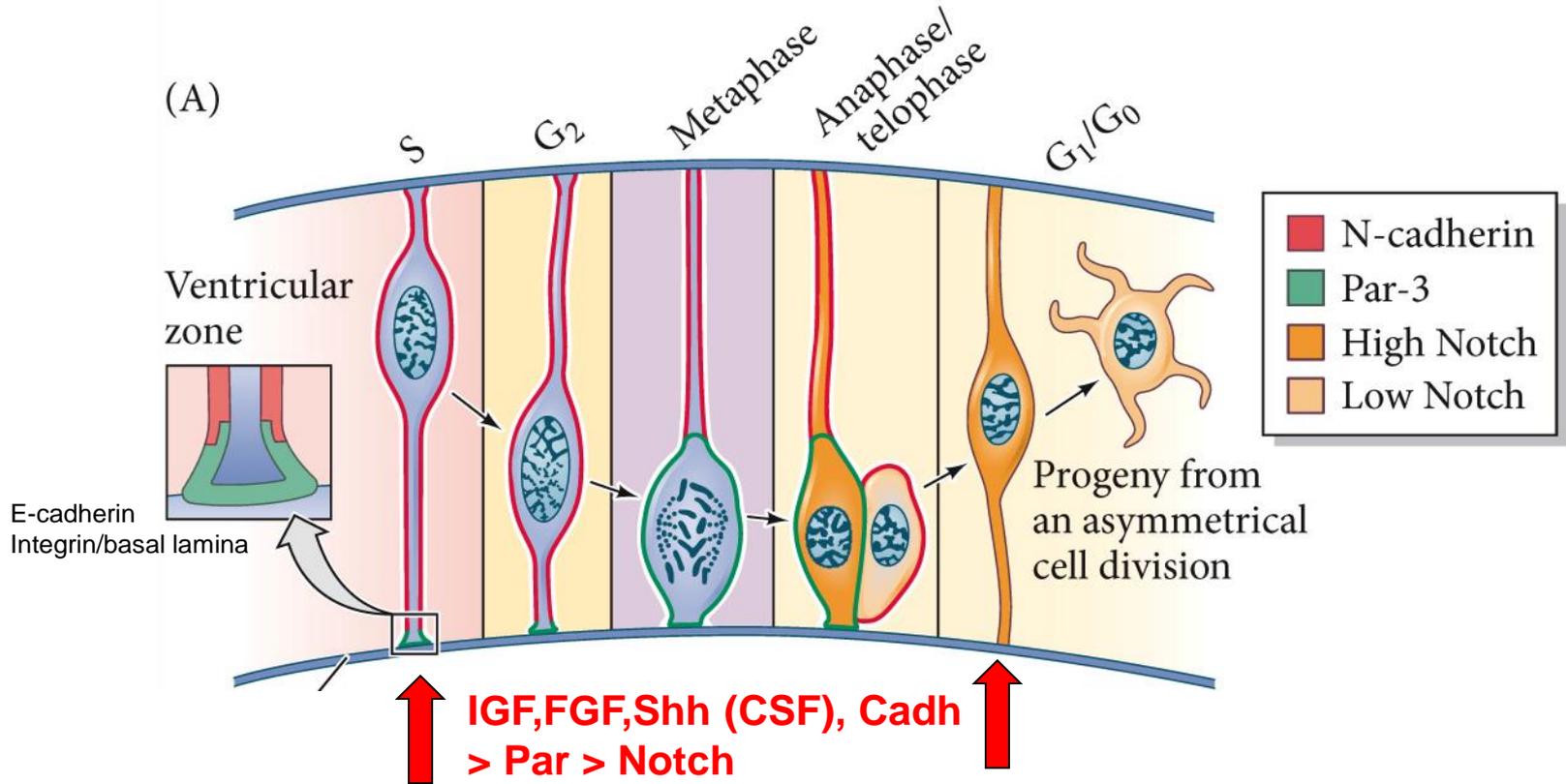


Figure 14.14 Asymmetrical division of radial glia mediated by Par3 and Notch.

Inheritance matters:

Old centriole + old luminal cilium = exposure to mitotic factors
 New centriole + new basal = no mitotic factors

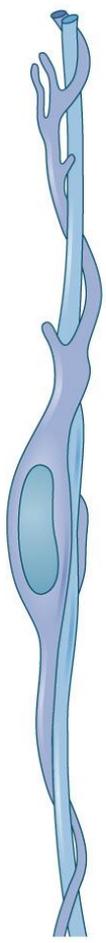
More Par-3 > high Notch, N-cadherin > stem cell
 Less Par-3 > Delta > neuron

Figure 14.11 NEC and later radial glia cells (RGC) provide tracks for neuron migration)

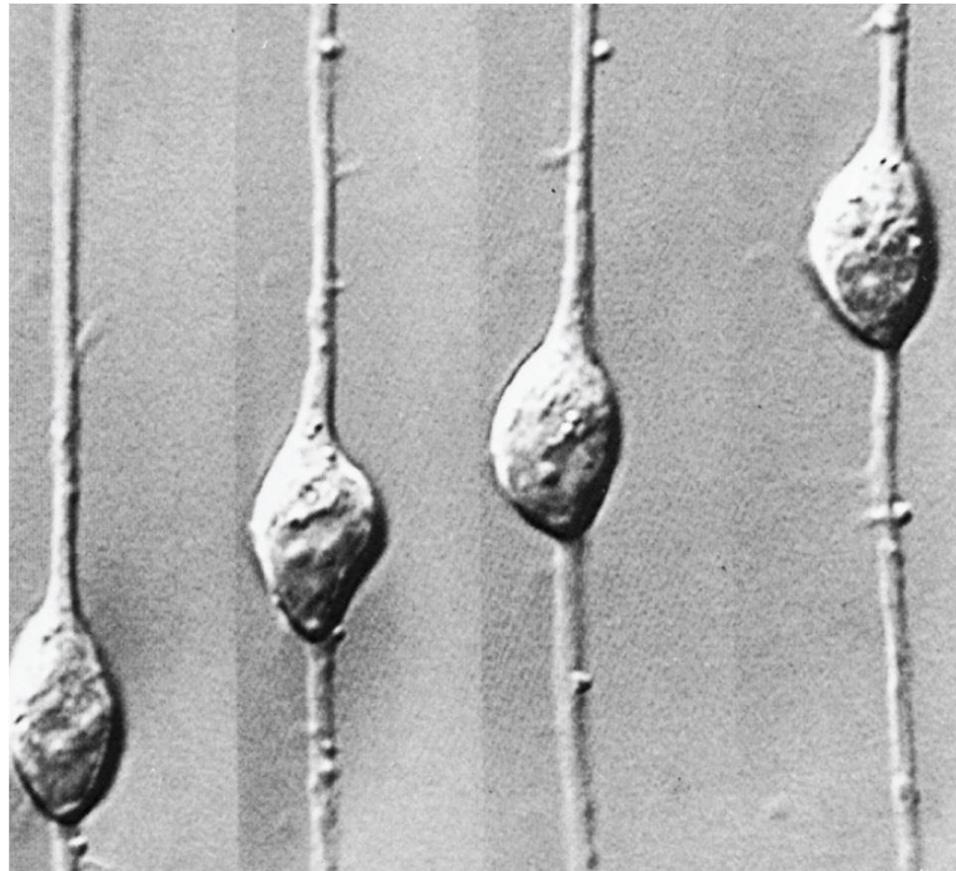
Neuron- epithelial cells

Neuron-Glia (astroactin, N-cadherins)

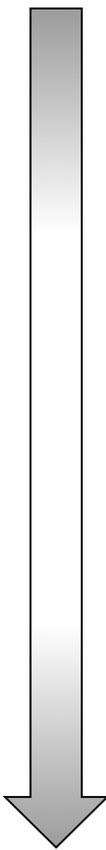
(A)



(B)

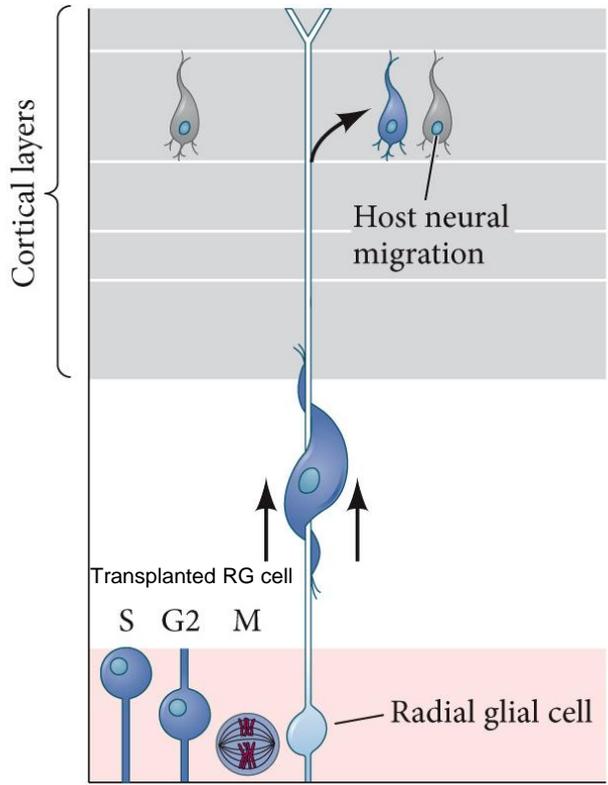


BDNF

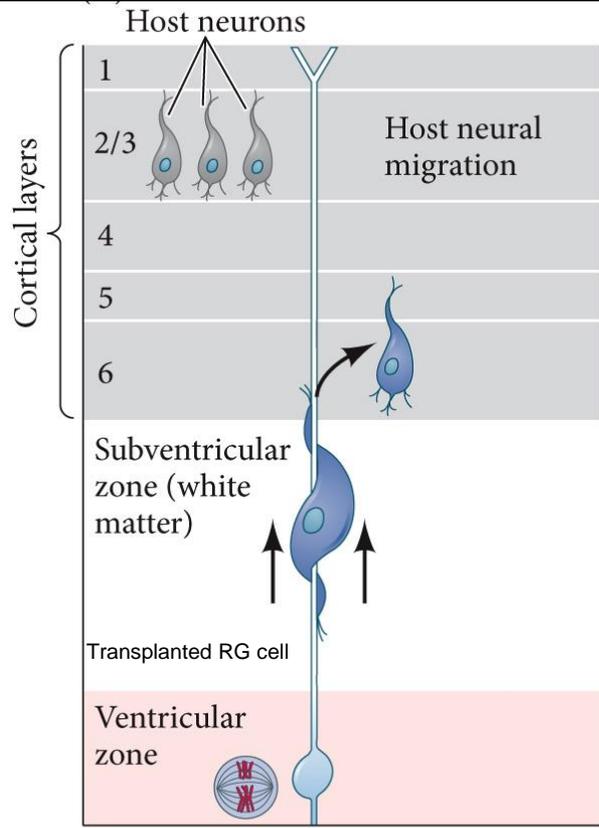
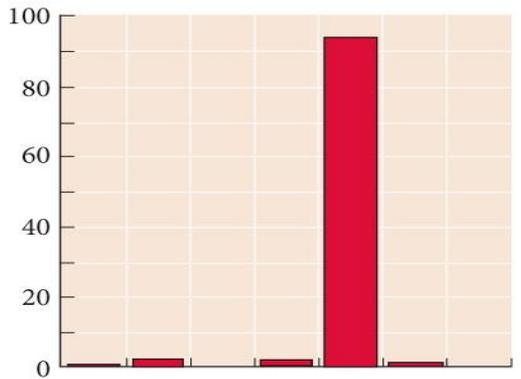


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Figure 14.12 Determination of cortical laminar identity in the ferret cerebrum (Part 3)



Host (conditional) fate when transplanted in S phase



Cell-autonomous fate when

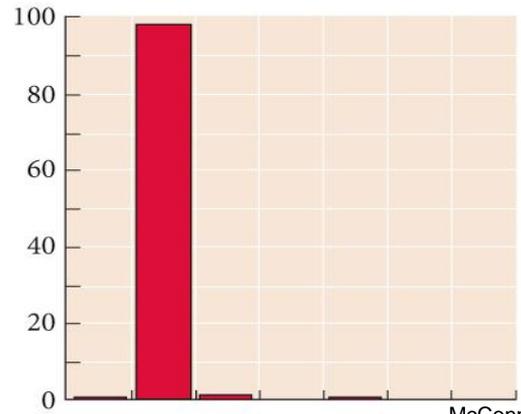
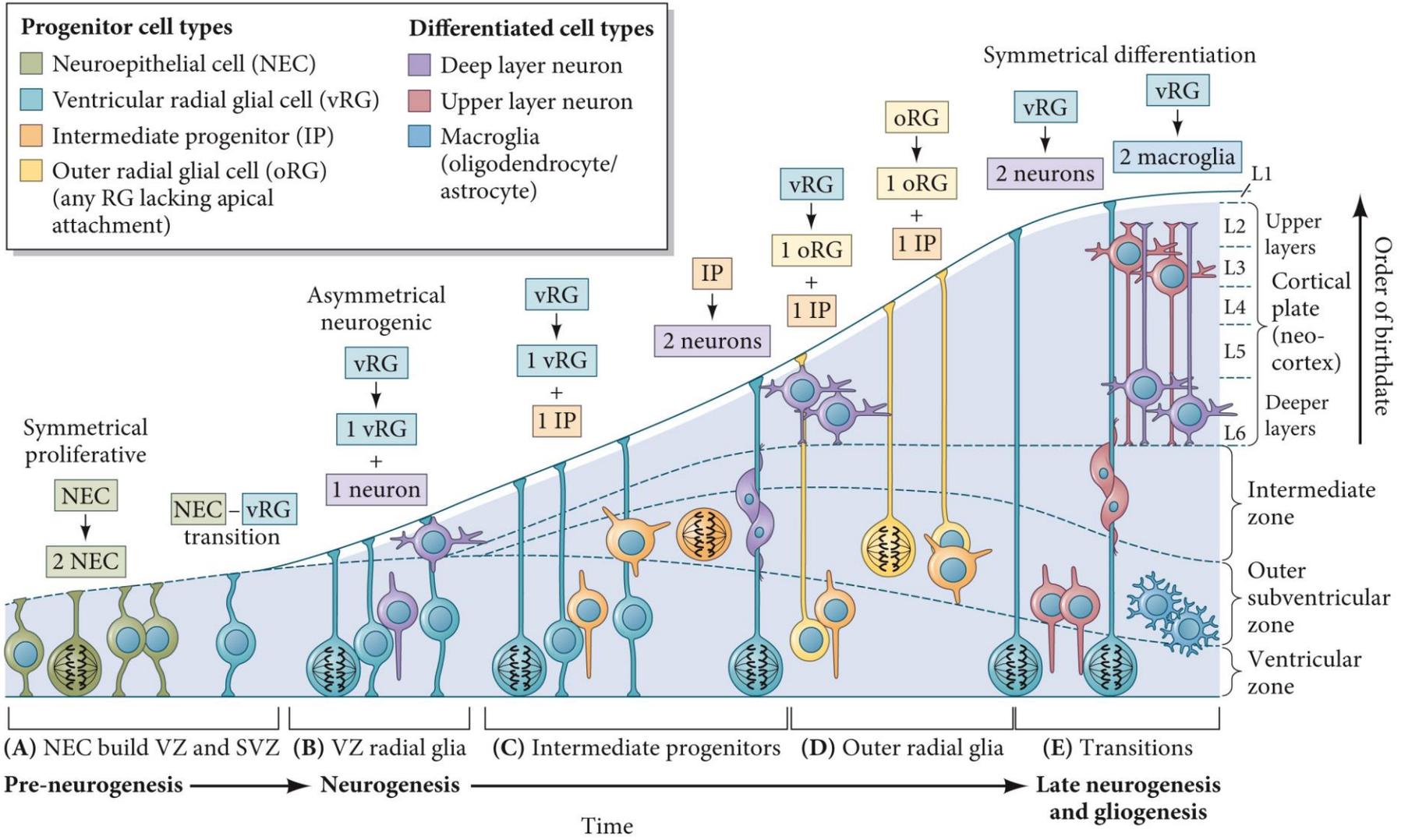


Figure 14.10 Summary model of neurogenesis in the cerebral cortex



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Figure 14.13 Building cortical layers by Caja-Retzius cells secreted *reelin* & *Disabled-1* signaling

(A)

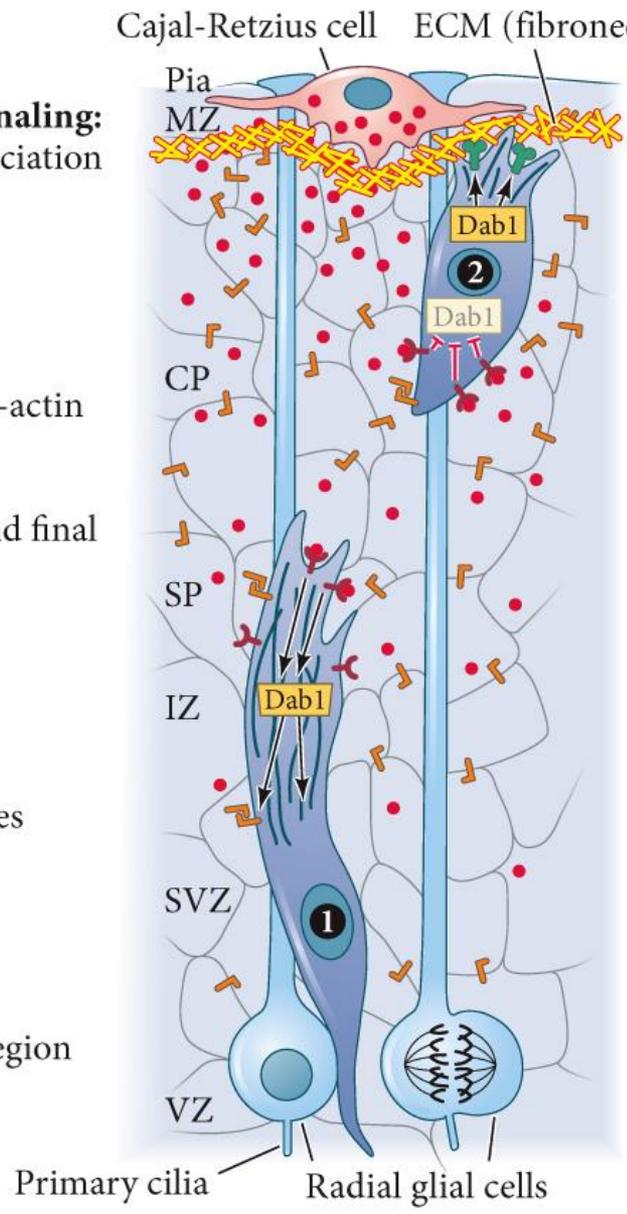
2 Reelin-Disabled-1 signaling:
Promotes integrin association with filopodial tip

High levels of Reelin inhibit Disabled-1 (negative feedback):
Leads to destabilized F-actin

Result:
Destabilized F-actin and final translocation

1 Moderate levels of Reelin activate Disabled-1 to:
Activates N-cadherin expression and stabilizes F-actin

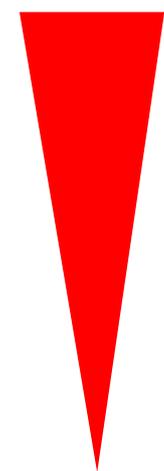
Result:
Promotes filopodial extensions and cell translocation toward region of highest N-cadherin



Attracting factors for gradients:

reelin

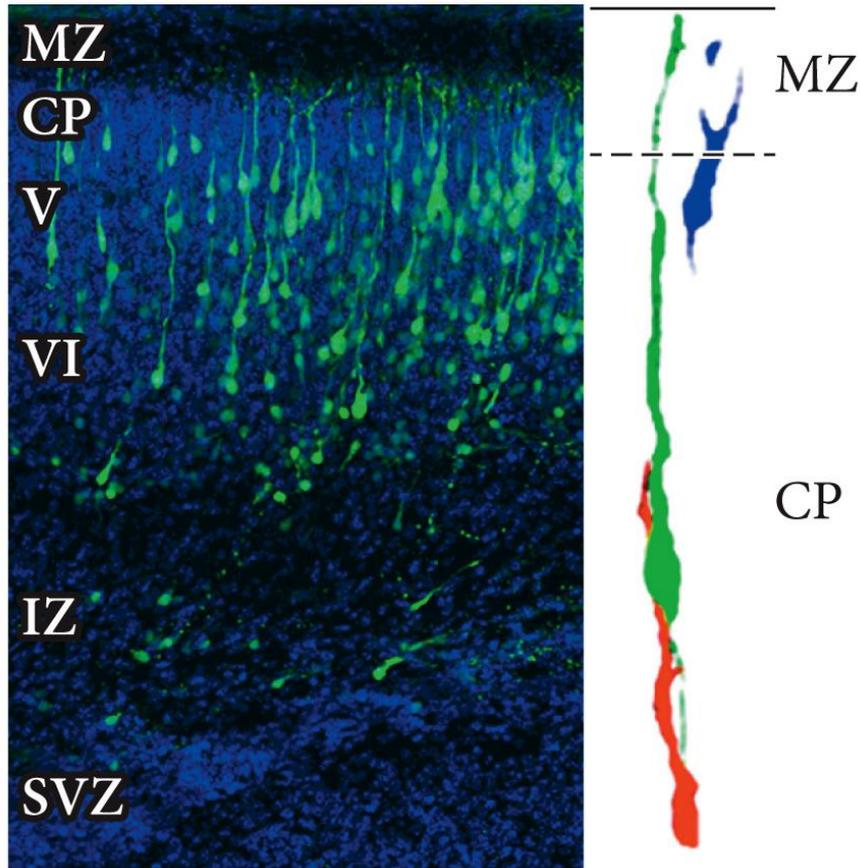
N-cadherin



Reelin ligand bound to its receptor	F-actin
Integrin receptors	N-cadherin
	ECM

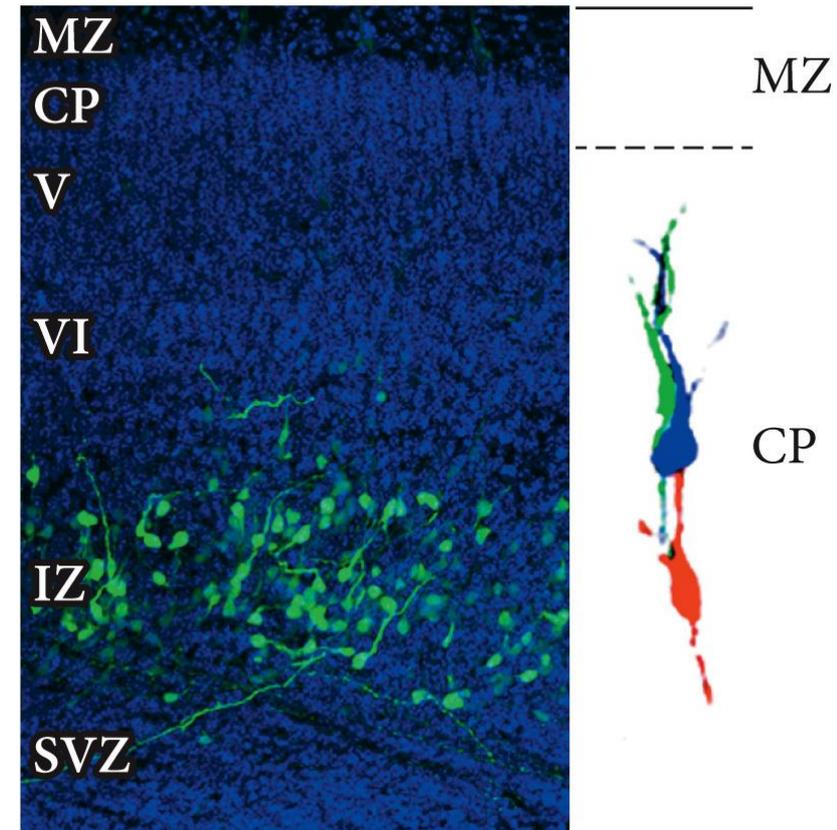
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(B) Wild-type



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(C) *Dab1* knockout
(only lost in green cells)

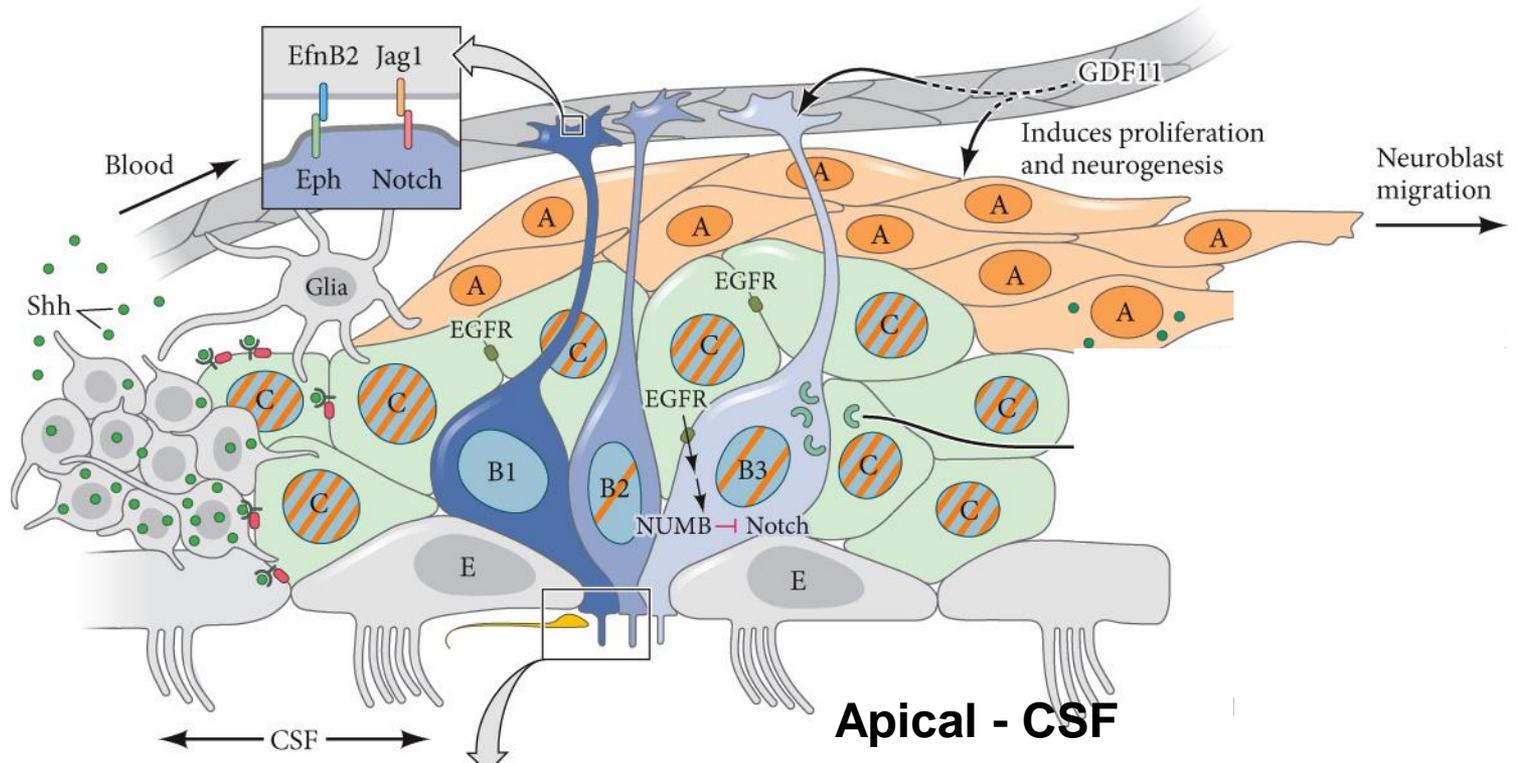


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GFP expressing progenitors

Figure 5.10 Modified Niche - ventricular-subventricular zone stem cell niche and its regulation

Basal
blood



Developmental progression of NSC (Radial Glial Cells):

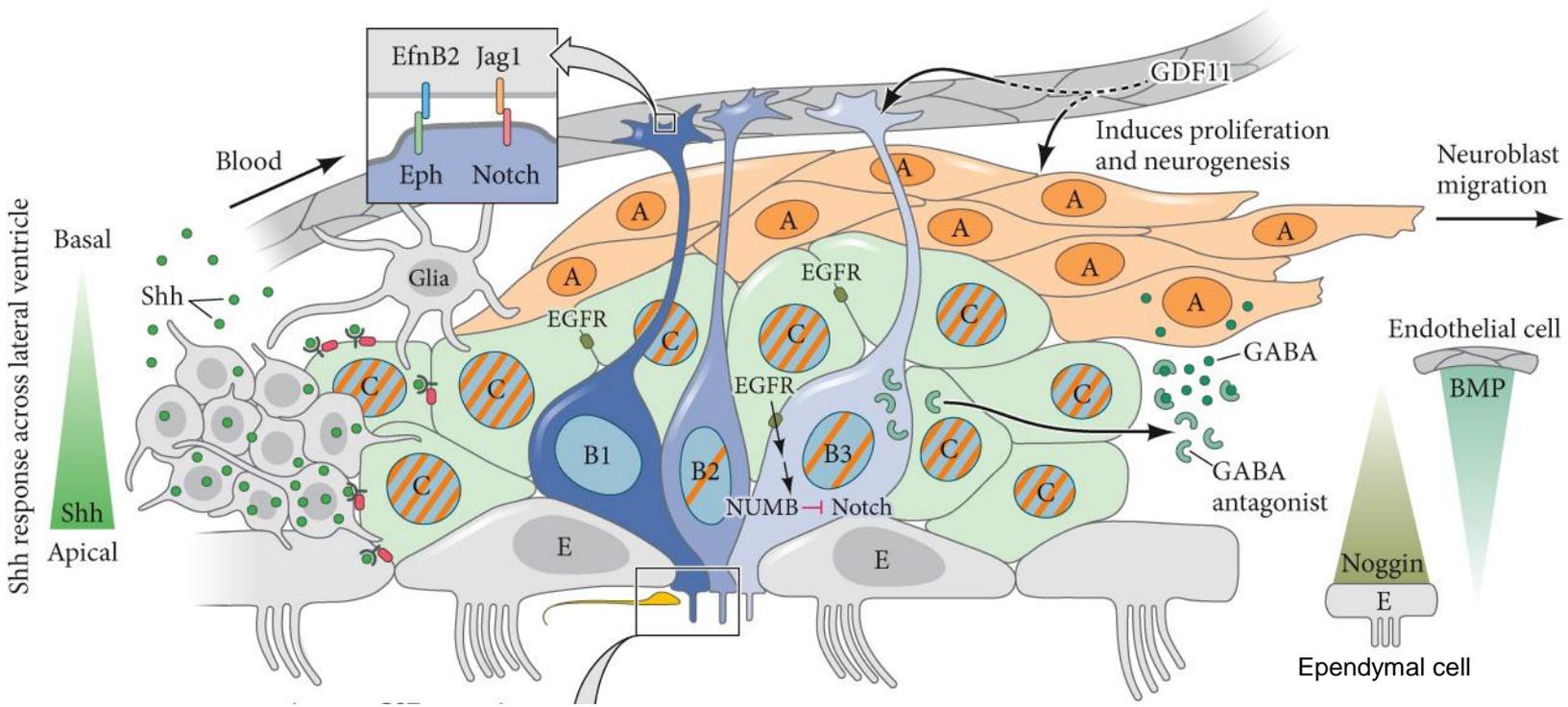
B1 quiescent (GFAP; (Notch-Asc-1) >

B2, B3 - activated proliferating (GFAP, Oscillating Notch-Asc-1; BIBP) >

C – proliferating Progenitor cells C (GFAP, Oscillating Notch-Asc-1, EGFR) >

A - migratory neuroblasts (Constant ASC-1> doublecortin)

Figure 5.10 Schematic of the ventricular-subventricular zone stem cell niche and its regulation

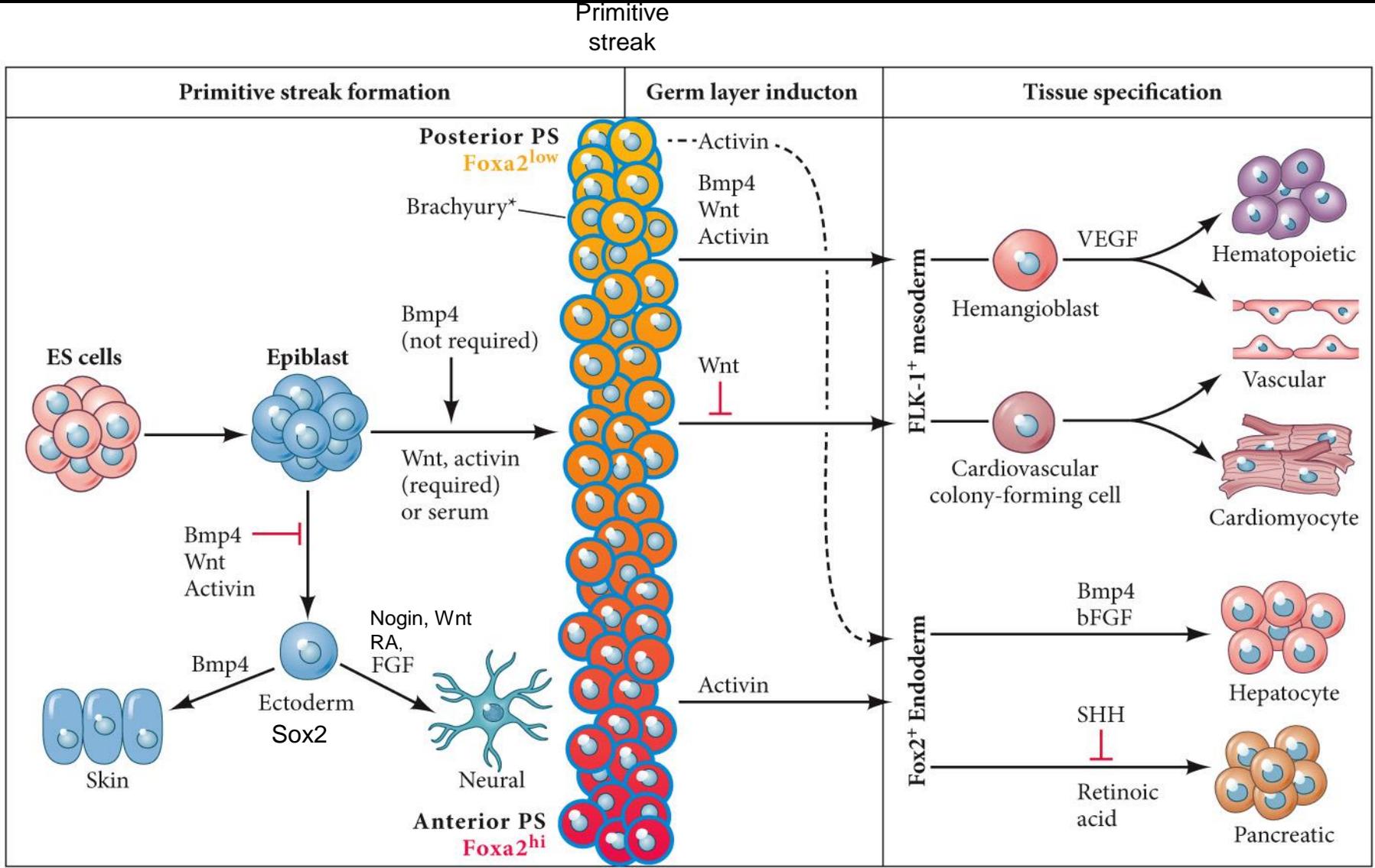


- Morphogenic gradients: BMP/Noggin; Shh
- Neuronal activity
- Bloodborne Growth Differentiation Factor (GDF11)
- Ependymal cell VCAM
- diminishing Notch signaling B1>B2>B3 (increasing proliferation) > A (off – differentiation)

STEM CELLS IN VITRO TECHNOLOGIES

workshop

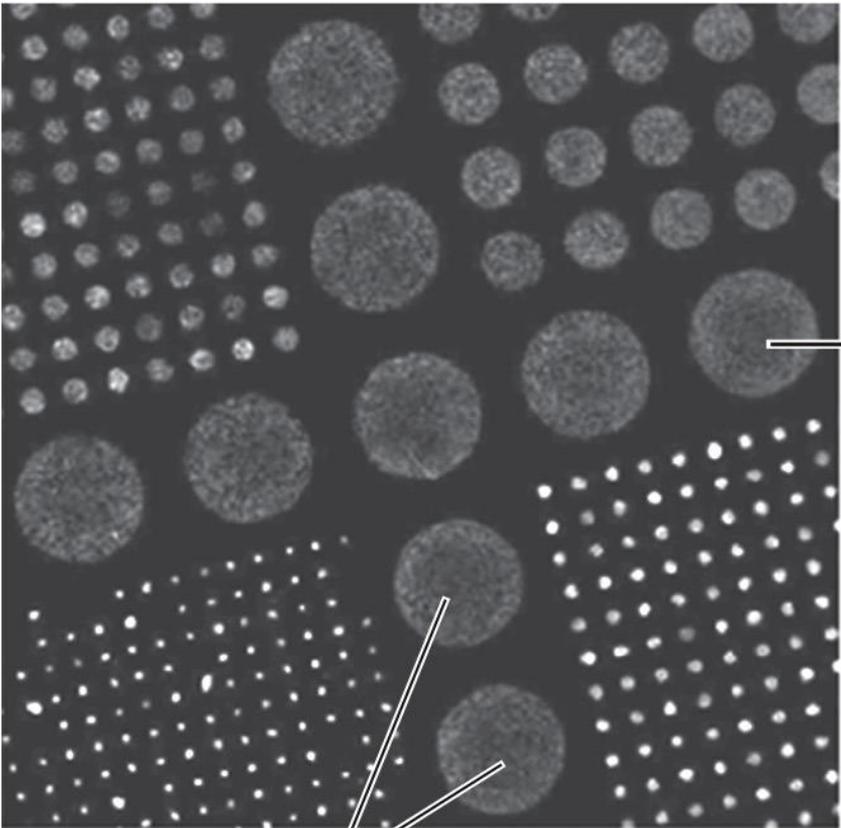
Figure 5.21 Inducing Experimental stem cell differentiation from ESCs using Paracrine & Transcription factors



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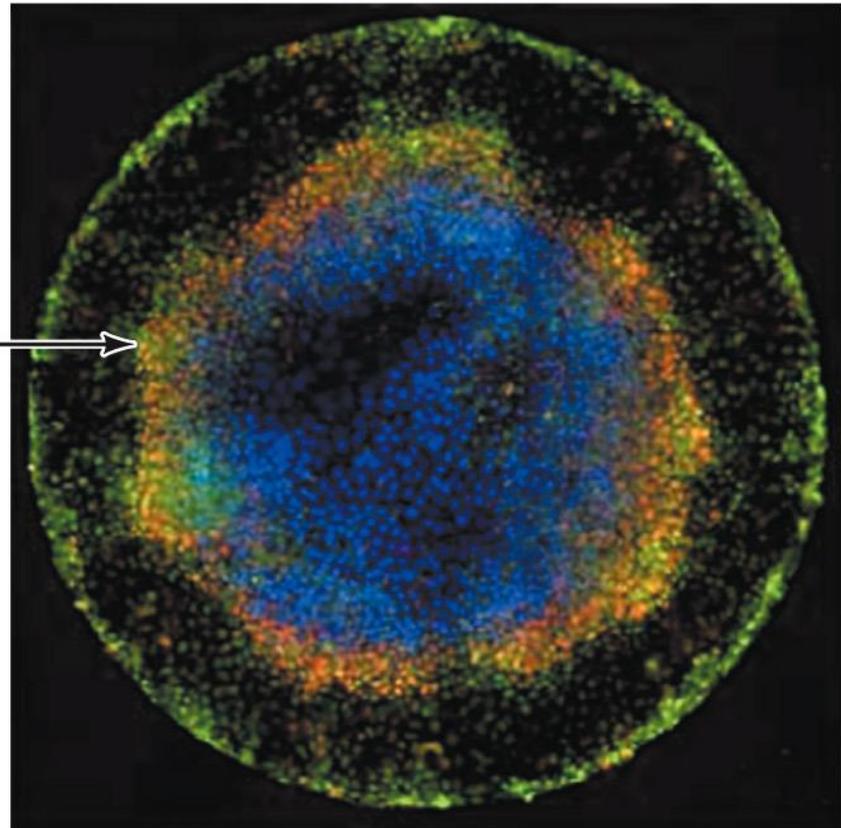
Figure 5.22 Human ESCs cultured in confined micropatterned discs demonstrate a pattern of differential gene expression similar to that seen in the early embryo

(A) Micropatterned cultures



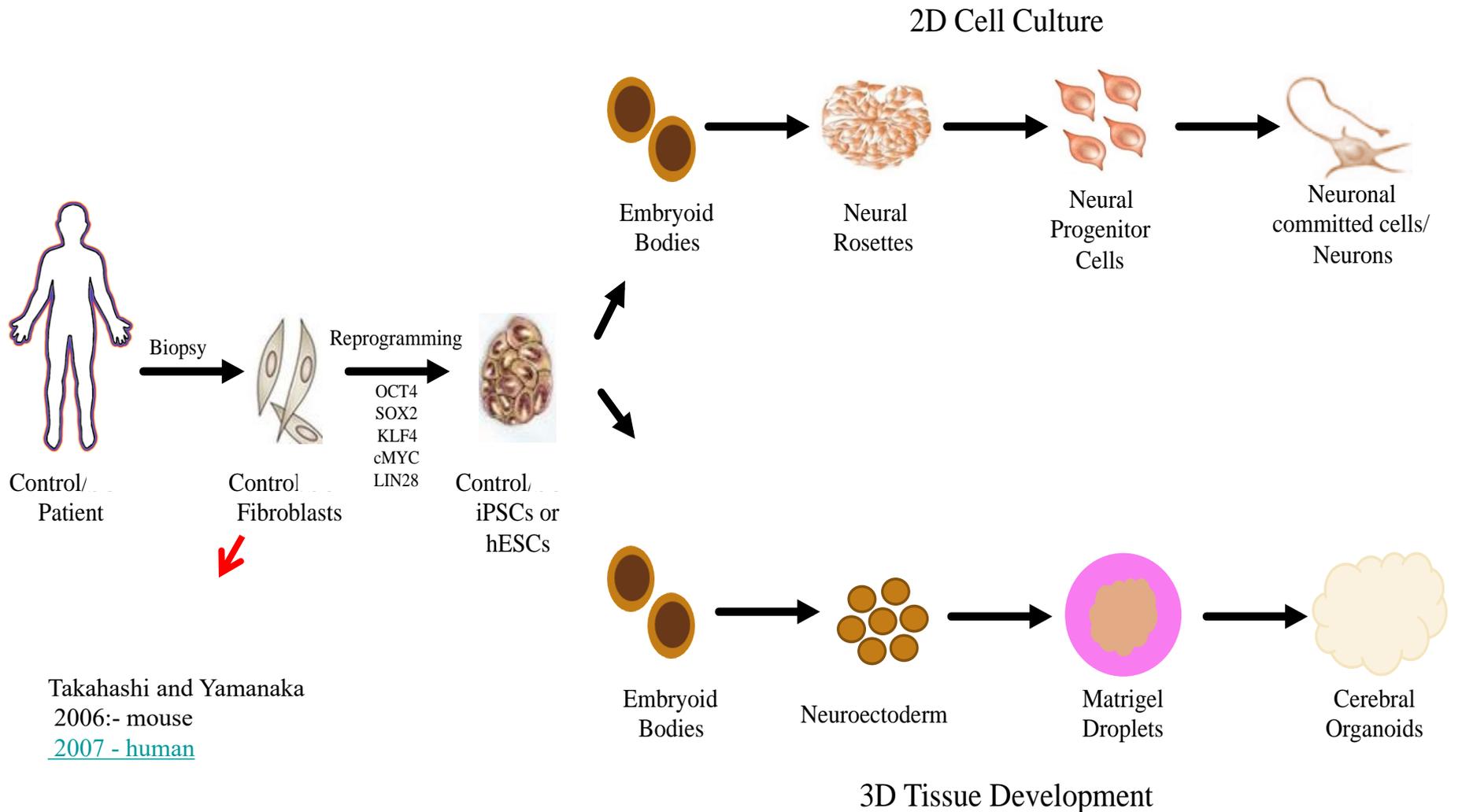
Micropatterned discs

(B) Radially patterned gene expression



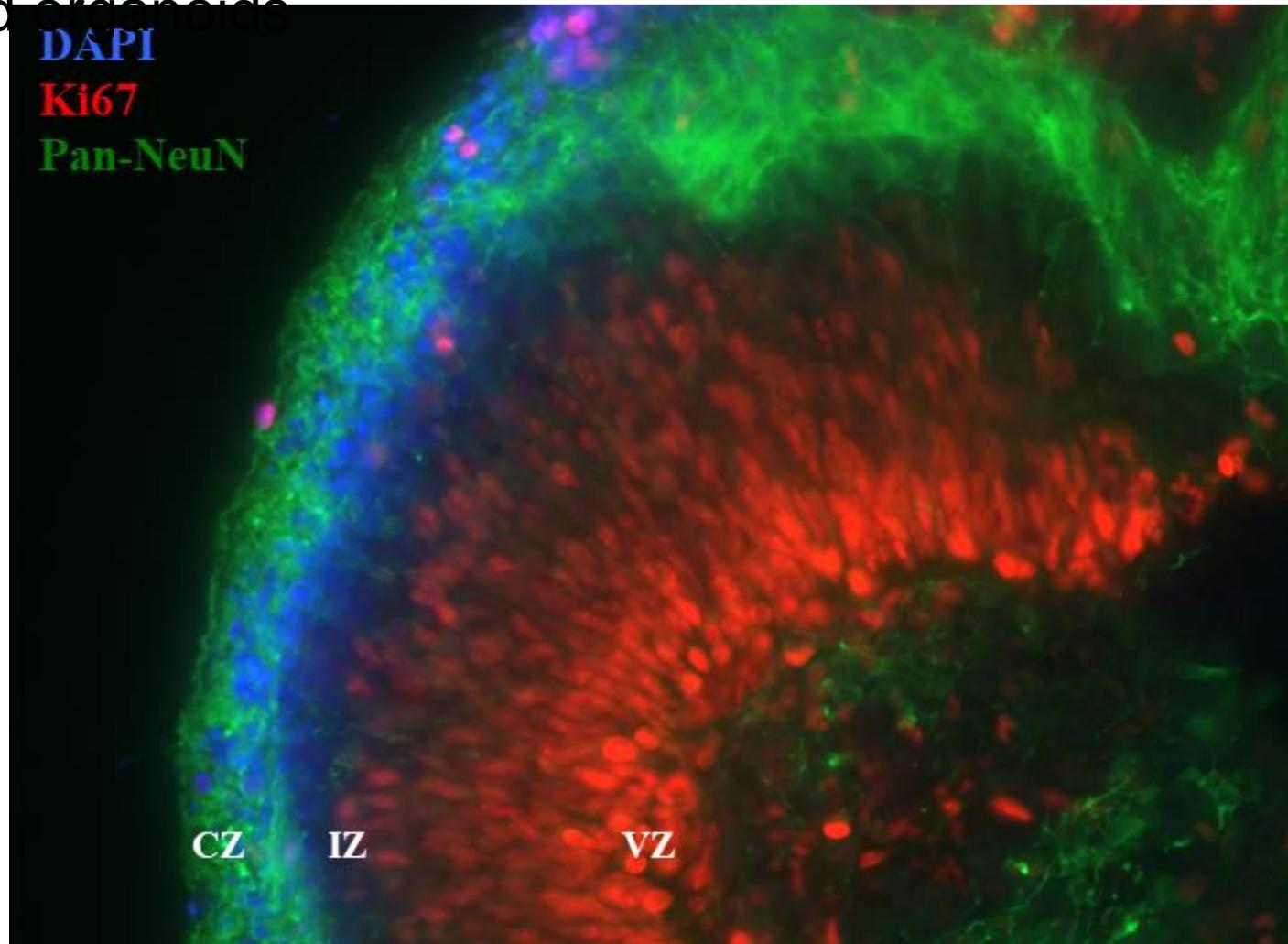
	Ectoderm (Sox2 ⁺)
	Trophectoderm (Cdx2 ⁺)
	Mesoderm (Bra ⁺)

Modeling neurodevelopment in vitro

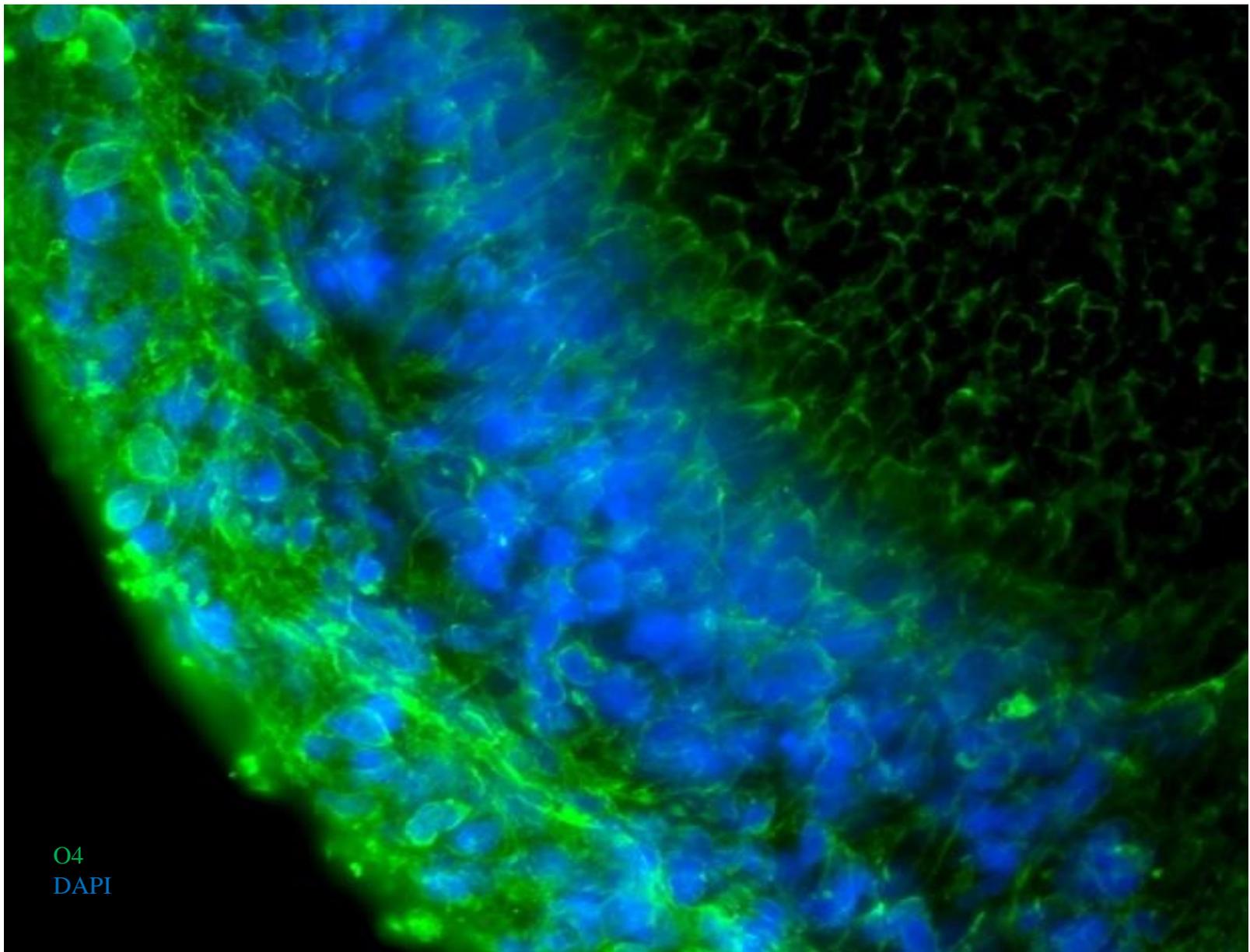


Apical to basal pattern of brain and organoid cortex development

14 day old organoids



Inner zone: **proliferating Ki67⁺ stem cells**; Intermediate zone:
Ki67⁺



22 day old organoids see **Olig4+** cells (marker for mature oligo)

MODELLING ABNORMAL BRAIN DEVELOPMENT WITH SCHIZOPHRENIA IPSC

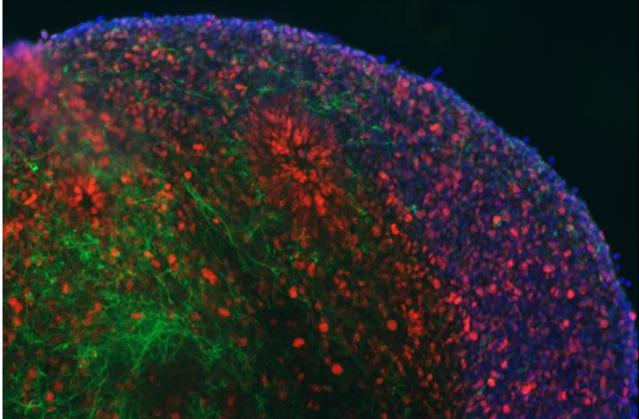
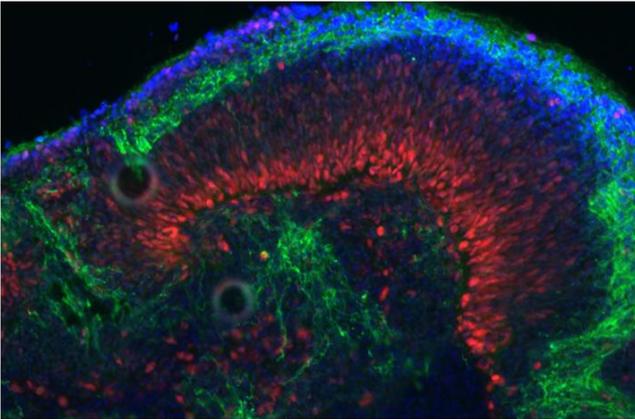
Ki67 PAN NEUN

Control

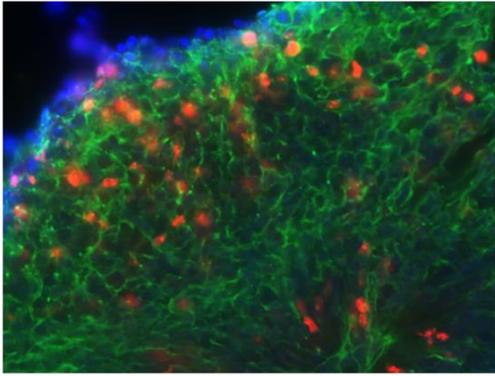
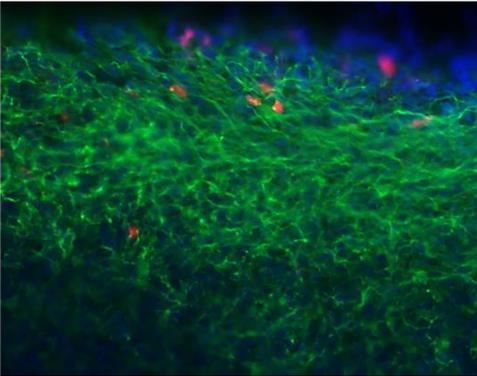
Schizophrenia

DAPI

2 wk

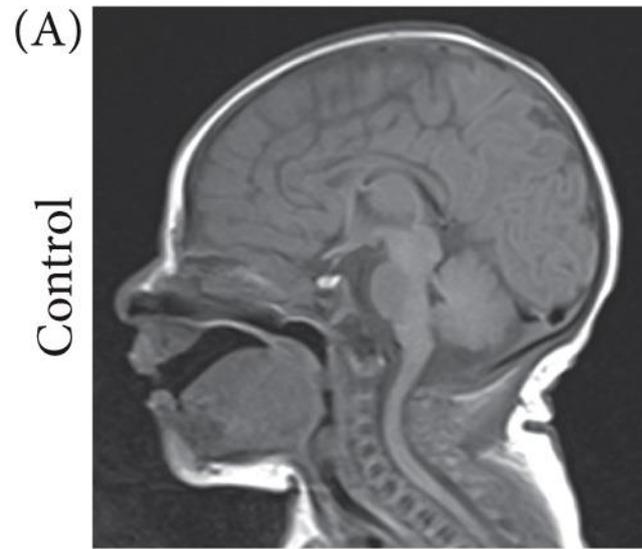


5wk



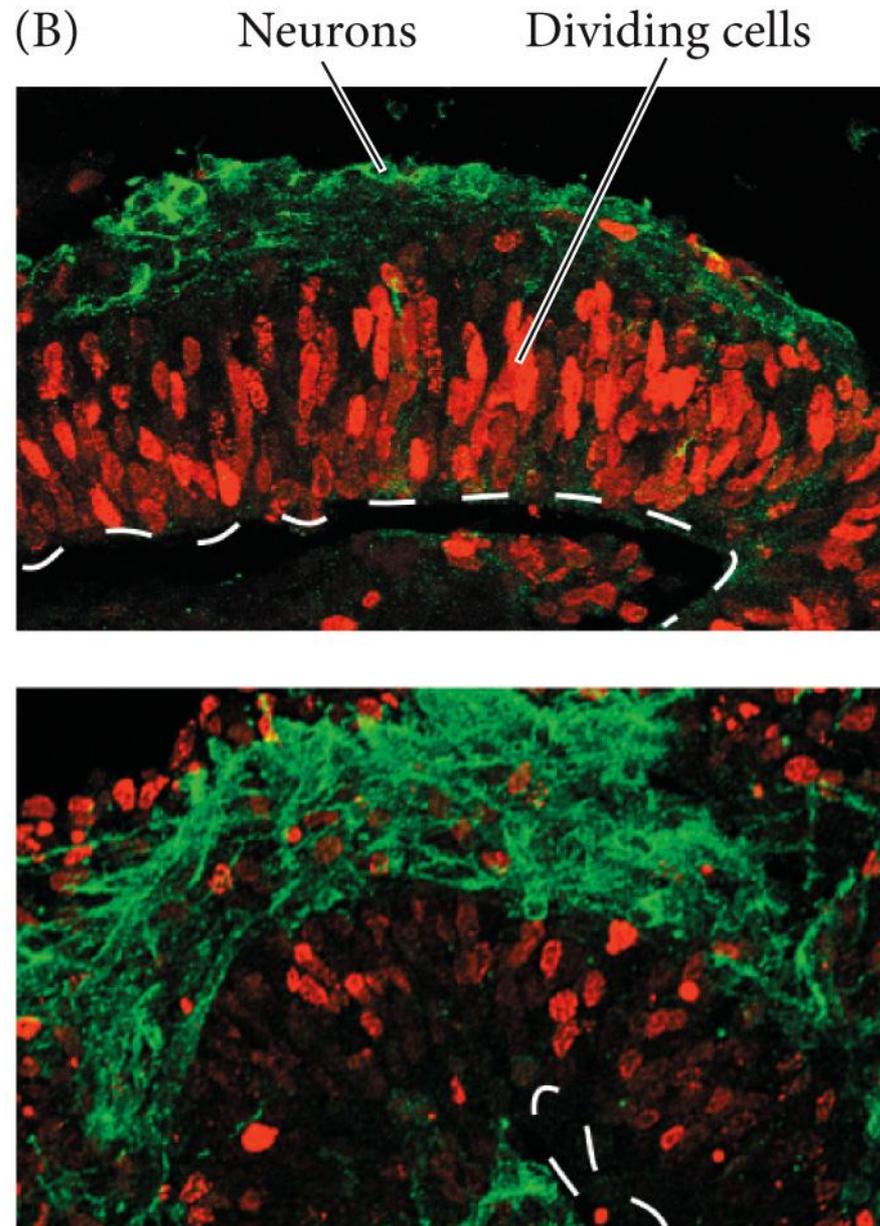
Stachowiak, C.A. Benson, Elahi, Narla, Freedman, Brennand, Klejbor, Stachowiak. Cerebral organoids reveal early cortical maldevelopment in schizophrenia – role of FGFR1. *Nature Translation Psych.*

Figure 5.27 Modeling human microcephaly with a patient-specific cerebral organoid (Part 1)



Microcephaly

Figure 5.27 Modeling human microcephaly with a patient-specific cerebral organoid (Part 2)



DEVELOPMENTAL BIOLOGY 11e, Figure 5.27 (Part 2)
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THE END